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# Effect of various temperatures on the nutritional compositions of fermented African locust bean (Parkia biglobosa) seed

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#### Abstract

The effect of temperature on the nutritional values of fermented African locust bean (*Parkia biglobosa*) seed were studied. Temperatures ranging between 40 to  $70^{0}$  C were used for the fermentation. Biochemical and physiological analysis were evaluated. Highest nutritional values were obtained with samples fermented at lower temperatures (40 and  $50^{0}$  C) and they all had acceptable end products while all the samples fermented with higher temperature (60 and  $70^{0}$  C) gave poor nutritional values with unacceptable end products. *Bacillus subtilis* was used as starter culture and fermentation was carried out for 5 days (120 hours). An increase in moisture, protein, crude fat contents and decrease in total carbohydrate and crude fibre were noticed for all the fermentation temperature variation. The Organic functional groups were identified and characterized using Fourier Transform Infrared (FTIR) spectroscopy. The effect of temperature on the morphological structure of fermented samples.

Keywords: Bacillus subtilis, fermentation, Iru, Parkia biglobosa, starter culture

#### 1. Introduction

Food fermentation is basically aimed at producing important nutrients or eliminating anti-nutrients [1]. This is necessary to improve the prevailing cases of malnutrition in Nigeria [2]. African locust bean seed was named *Parkia biglobosa* after a Scottish botanist and surgeon called Mungo Park [3]. *Iru* is a product of alkaline fermentation of *Parkia biglobosa* seeds which is popularly used as food seasoning and condiment in West African especially Nigeria. It is a good source of protein supplement for a protein deficient country and low income earners who could not afford animal protein.

*Parkia biglobosa* tree has application in both medicine and food additives to the indigenous people of Africa. The tree serves various medicinal purposes such as cure of diarrhea and hypertension since it contains a deposit of histamine which dilates the blood vessels and allows free flow of blood <sup>[4, 5]</sup>. <sup>[6] [7]</sup> reported fermented African locust bean seeds to be rich in

protein with 39-47%, 31-40% of fat or lipid and 12-16% of carbohydrate. Published studies identified *Bacillus* spp. as the main microorganism involved in the fermentation of most legumes especially *Parkia biglobosa* with *Bacillus subtilis* as the predominant microorganism [7, 8, 8, 10, 11].

Fermentation of African locust bean seeds is a traditional art that is practiced in rural areas with rudimentary utensils which did not put hygiene into consideration. Since there is no standard method for the fermentation, every producer produces base on their culture and traditions which brings variation in the quality of *Iru* [12]. The length of production and also the methods of fermentation vary from one producer to another and with the final intended use, resulting to non

uniformity in products <sup>[13]</sup>. Flavour is considered the quality index of fermented African locust bean and plays an important role in consumer acceptability <sup>[14]</sup>. Although fermented African oil bean seeds have typical flavour and appealing organoleptic quality, differences in flavour range and intensities exist. These vary perhaps as attributes of producer organisms which are due to the various volatile compounds produced by the fermenting population <sup>[15]</sup>, this study used FTIR to study the variation in the volatile compounds as a result of change in temperature.

This study was carried out to know the effect of temperature variation on the nutritional composition and to bring out the optimum temperature that would yield the best nutrient composition for the fermentation of *Iru*.

#### 2. Materials and Methods

Parkia biglobosa seeds used were purchased from local market.

All the chemicals used were of analytical grade from Sigma manufacturing industry. The starter culture used was prepared in Covenant University Microbiology laboratory, Sango-Ota, Nigeria.

#### 2.1 Laboratory preparation of Parkia biglobosa

The *Parkia biglobosa* purchased from the market were processed using method of <sup>[4]</sup>.

# 2.2 Preparation of Bacillus subtilis

Preparation of the inoculum used was carried out using method of [12].

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#### 2.3 Inoculation of Seeds

200 g of processed seed was inoculated with freshly prepared *Bacillus subtilis*. Five flasks labelled 24 hours to 120 hours were placed in a thermostatically controlled fermenter. At the end of every 24 hours samples were removed from the fermenter and packed into an air tight container which is stored in the freezer for further analysis.

### 3. Biochemical Analysis

# 3.1 Proximate Analysis of Fermented African Locust Bean

**Seeds:** The parameters determined were % moisture, % crude protein, % fat, % ash, % crude fibre and % total carbohydrate compositions. These were determined using the [16] method.

# 3.2 Temperature Monitoring

This was done using the temperature probe fabricated with the fermenter used for the fermentation process.

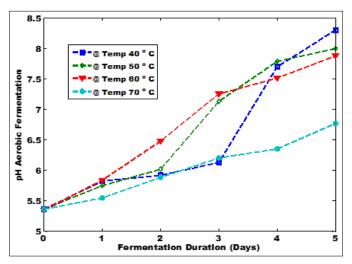
#### 3.3 Physiological Analysis

This is majorly the physical characteristics of the condiments such as taste, aroma, colour, texture and appearance.

#### 4. Results & Discussion

The seed was fermented to a protein based vegetable condiment identified by its appearance, aroma, and taste. The smell was prominent after the third day (72 hours) of fermentation. The substrate became dark and soft with characteristics aroma similar to ammonia.

#### 4.1 The pH



**Fig 1:** Effect various temperature on the pH of the fermented samples.

Prior literature works confirmed that the fermentation of African locust bean (*parkia biglobosa*) seed to Iru is an alkaline fermentation [12, 17, 18].

Figure 1 showed that lower temperature favoured increase in pH value towards alkalinity during fermentation. This is due to the ability of *Bacillus subtilis* to produce protease to hydrolyse protein and obtain amino acids and ammonia as source of carbon and energy for growth, respectively. Alkaline conditions provided a suitable environment for the hydrolysis of proteins by *Bacillus subtilis* which resulted in the

production of amino acids and the release of ammonia which is alkaline in nature [14].

# **4.2** The Proximate Analysis of African Locust Bean Seeds

Figures 2a-5h shows the effect of temperation variation on the Proximate analysis of African locust bean seeds.

#### 4.2.1 % Moisture content

The moisture content increased with fermentation duration in all the temperature [40-70°C]. Analysis showed that the initial moisture content was 68 % which later increased to 76 % after the third day of fermentation. The raw unfermented seeds had 9.62 %, while raw cooked unfermented seeds had 56.33 % moisture content. During fermentation the higher the fermentation temperature, the higher the moisture content. The increase was probably due to the water used in subsequent processing such as soaking, dehulling and boiling of the raw seeds prior to fermentation processes and the increase in temperature which probably generated steam, since the fermenting vessels were tightly covered. [19, 20] reported that the metabolic activities of some microorganisms during fermentation gives out moisture as one of their end products. Increase in moisture content was noticed in all the fermentation temperature conditions.

#### 4.2.2 % Ash content

Figures 2a-5h shows that ash content decreased with fermentation duration (days) and increased with fermentation temperature. Increase in ash content was noticed in fermentation temperature 40 and 50°C while a decrease was also noticed at 60 and 70°C. The ash content varied between 2.6-4.0 %. The decrease in percentage ash content was as a result of long hours of boiling, soaking in water and dehulling. A30-40% reduction was noticed, which implies that the total mineral content of the seeds resides in the hull of the seeds which are leached during processing [13, 19]. The reduction in percentage ash content may also be due to the utilization of some essential salts during fermentation by microorganisms for their metabolic activities.

# 4.2.3 % Crude protein

Several research works have been carried out on how fermentation enhances protein value of Parkia biglobosa seeds, this was established in figure 2a with increase in protein content at the third of fermentation. The protein composition decreases as temperatures increases. Figures below showed that at every fermentation temperature protein content increased from the first day up to the third day and declined from the fourth day. The % yield of protein from the fermentation of Parkia biglobosa at any given day of fermentation declines with increase in fermentation temperature from 40 to 70 °C. The protein yield at 40 °C on the third day of fermentation is 52.7 %, it reduced to 25.6 % on the third day at a temperature of 70 °C. This is shows that bacillus subtilis functions well at lower temperature (40-50 <sup>0</sup> C). Thus maximum yield of protein is achieved at the third day of fermentation with temperature 40 °C.

#### **4.2.4 % Crude fibre**

Fermentation reduced the percentage crude fibre of the

substrate (fermenting seed) with days of fermentation. An increase in fermentation temperature led to decrease in the percentage crude fibre. Temperature seems to have minimal or no impact in the Crude fibre of African locust bean seeds during fermentation. The patterns in figures 2a-5h reduced progressively, showing that there is a consistent decrease in the percentage crude fibre content.

#### 4.2.5 % Fat

Proximate analysis of both raw and cooked African locust bean seed gave about 17.33 % and 19.33 % of fat content respectively. Increase in fermentation temperature led to a decrease in % fat content. There was an increase in the % fat content with days of fermentation. The activity of lipolytic enzyme is faster with a slight increase in temperature during fermentation process but an elevated temperature above the optimum operating temperature of the fermenting microorganism- *bacillus subtilis* which is 40-50 °C will either make the organism go dormant and reactivated with a favourable conditions or die.

#### 4.2.6 % Total Carbohydrate (Cho)

A decrease in total carbohydrate which is expected since it is the substrate that is been consumed by the fermenting microorganisms. The loss in carbohydrate during soaking and boiling is attributed to the leaching of soluble carbohydrates like sugars into the cooking water. Loss in carbohydrate during fermentation may also be as a result of the fermenting organisms that utilize some of the sugar for growth and metabolic activities [21]. The higher the temperature the lower the reaction since the fermenting microorganism cannot function well at high temperature, hence low conversion of carbohydrate to sugar.

In summary, the highest compositions were obtained on the third day of fermentation at temperature 40 °C, this gives the optimum operating temperature for the fermentation of African locust bean seeds. This is in support of the claims in literature that the fermenting microorgansim in African locust bean seeds has an optimum temperature with minimum value of 18 and a maximum value of 45 °C.

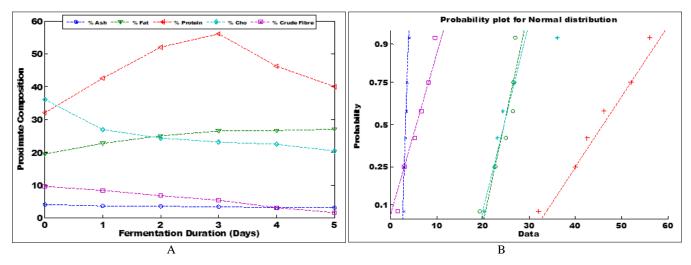


Fig 2a&b: proximate analysis at 40o C

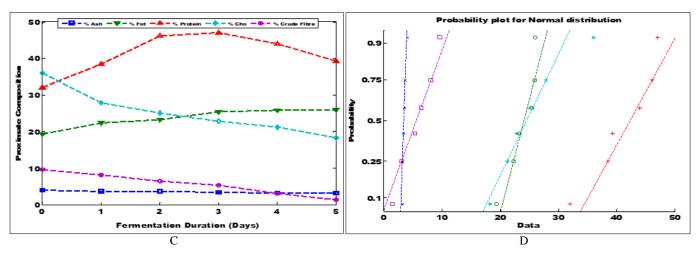


Fig 3c&d: proximate analysis at 50 ° C

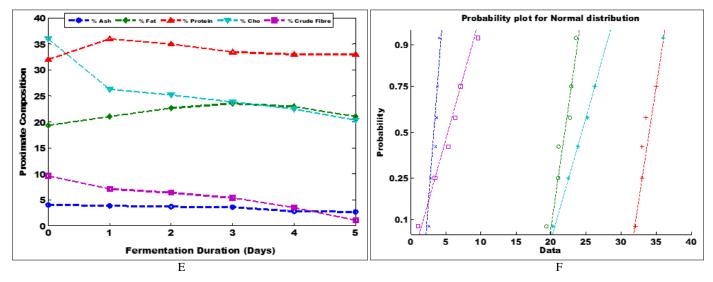


Fig 4e&f: proximate analysis at 60 ° C

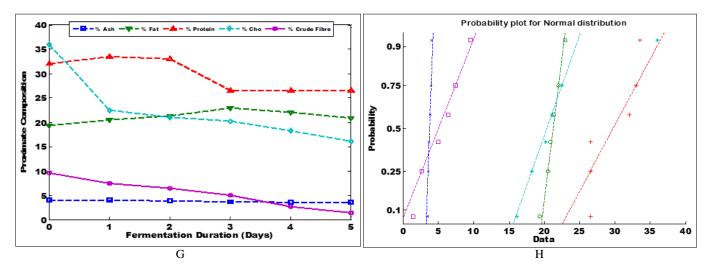


Fig 5g&h: proximate analysis at 70 o C

The normal distributions of each components show that at 40 °C, the probability of obtaining crude fibre, ash, cho, fat and protein is 0.25, 0.25, 0.6, 0.6 and >0.9 (Figure 2b). Hence, the probability of distribution is proportional to the proximate composition (Figure 2a).

The normal distributions of each components show that at 50 °C, the probability of obtaining crude fibre, ash, cho, fat and protein is 0.25, 0.25, 0.5, 0.5 and >0.8 (Figure 3d). Hence, at an increased temperature, the probability of distribution for cho, fat and protein decreased while crude fibre and ash remained constant (Figure 2a). Hence, at higher temperature, the probability of distribution in composite-crude fibre and ash is not proportional to the proximate composition (Figure 3c).

The normal distributions of each components show that at 60 °C, the probability of obtaining crude fibre, ash, Cho, fat and protein is 0.15, 0.15, >0.1, >0.1 and >0.9 (Figure 4e). Hence, the probability of distribution is proportional to the proximate composition (Figure 4f).

The normal distributions of each components show that at 70 °C, the probability of obtaining crude fibre, ash, Cho, fat and protein is 0.40, 0.40, 0.7, 0.7 and >0.9 (Figure 2b). Hence, at a

specific temperature, the probability of distribution is inversely proportional to the proximate composition (Figure 2a).

# 5. Physiological/Organoleptic Analysis

Panel of 10 assessors were used with the following results;

Table 1: Physiological/Organoleptic Analysis of the Final Product

|                          | Colour | Taste | Texture | Appearance |
|--------------------------|--------|-------|---------|------------|
| Dislike Extremely        | None   | None  | None    | None       |
| Dislike Slightly         | None   | 10%   | None    | None       |
| Neither Like Nor Dislike | 20%    | None  | 10%     | None       |
| Like Moderately          | 80%    | 90%   | 70%     | 70%        |
| Like Extremely           | 90%    | 90%   | 90%     | 90%        |

Purchase Interest 100% while Overall Acceptability of 90% were obtained.

Only the sample fermented for 72 hours with fermentation temperature  $40^{0}$  C was acceptable with respect to taste, colour, appearance and aroma, this shows the optimum fermentation conditions for the fermentation of African locust bean seed.

# 6. Identification of Organic Functional Groups Present in African Locust Bean (*Parkia biglobosa*) Seeds.

The Organic functional groups were identified and characterized using Fourier Transform Infrared (FTIR) spectroscopy (Bruker VideoMVP<sup>TM</sup> Single Reflection ATR Microsampler Spectrometer model). This is an accurate and effective way of determining the absence or presence of different functional groups in a molecule.

The spectrum in raw unfermented African locust bean has a characteristics strong stretching broad bond of O-H, hydrogen-bonded Alcohol, Phenols and carboxylic acids compound, with absorption frequency range of 3271.68 cm<sup>-1</sup>. A stretching vibration of C-H, Alkynes with strong and sharp intensity is observed at the same absorption frequency. Similarly, a stretching broad N-H medium but secondary band amine was also noticed. Bands such as stretching frequency of alkane C-H and alkene =C-H with strong and medium intensity was noticed at absorption frequency of 2924.52 cm<sup>-1</sup> respectively. All sample analysed shows the same properties as above but at different characteristics absorption of frequency range. This explains the similarity in aroma (odour) during fermentation at various temperature. The intensity of each compounds found at different stages of fermentation temperature differentiates them from one another, this accounts for the increase in aroma (odour) during and after fermentation.

C-Cl and C-Br (Alkyl halides and aromatics compounds) are the compound which was observed only on the 5<sup>th</sup> day of fermentation at temperature 50 °C. This probably might be responsible for the strong undesirable odour perceived after the fourth day of fermentation at higher temperatures (60-70°C).

# 7. Morphological Structure

The Morphological structures of African Locust Bean (*Parkia biglobosa*) Seeds viewed using SEM (Scanning Electron Microscope) revealed the following;

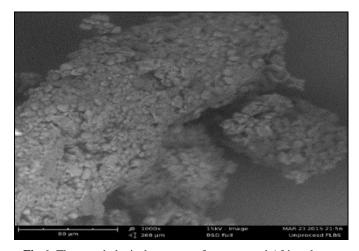


Fig 6: The morphological structure of unprocessed African locust bean seeds

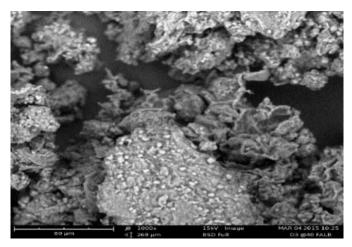


Fig 7: The morphological structure of fermented sample on the third day at 40°C

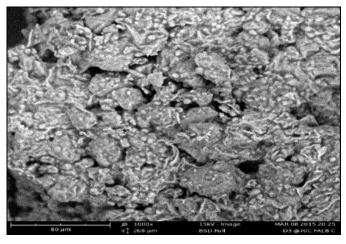


Fig 8: The morphological structure of fermented sample on the third day at 70°C

The result obtained from SEM showed the effects of temperature and fermentation on the morphological structure of both raw and fermented sample. This revealed the differences in processing and fermentation stages. Figure 6 revealed the morphological structure of the unprocessed seed with an agglomerated cohering image with a coarse and wrinkled corrugated surface (No reaction or fermentation observed). Figure 7 revealed the morphological structure of sample fermented for three days at temperature 40°C. A wider agglomerated and non-cohering structure with a wider pores compared to figure 6, this was attributed to the introduction of heat. This confirmed that compounds were been broken down into smaller units such as carbohydrates. Granules were also still visible, probably protein but adhere more to the surface of the structure (Reaction observed).

Figure 8 revealed the morphological structure of the third day fermented sample at fermentation temperature  $70^{\circ}\text{C}$ .

Agglomerated with a cohering structure was seen. Since the structure were compact and dense, disintegration is probably to be low or absent, hence little or no reaction.

This is probably the reason why all the products at this fermentation temperature were not acceptable. Narrower pores

with more fissures were also noticed all over the surface. [22, 23] reported that the morphology of fermented and unfermented African locust bean seeds depends on the botanical sources of starch.

#### 8. Conclusion

The highest nutritional composition of fermented  $Parkia\ Biglobosa$  was obtained at fermentation temperature  $40^{\circ}$  C on the third day (72 hours). Elevation in the fermentation temperature denatures nutrient and the nutritional values. The higher the temperature, the lower the nutritional values in the fermented seed.

This work discovered that fermented *Parkia biglobosa* was discovered to be very rich in protein (about 52%), increase in its consumption will reduce the risk of nutrient deficiencies in consumers.

**9. Conflict of Interests**: The authors declare that there is no conflict of interests regarding the publication of this paper.

#### 10. References

- Liman AA, Egwin P, Vunchi MA, Ayansi C. Lipase Activity in Fermented Oil Seeds of Africa Locust Bean, (*Parkia Biglobosa*), Castor Seeds (*Ricinu Communis*) and African Oil Bean (*Pentaclethra Macrophylla*). Nig J Bas & App Sc., 2012; 18:136-140.
- Steinkraus KH. Indigenous fermented foods involving and alkaline fermentation. Food Res Int, 1995; 54:234-237
- 3. Sadiku OA. Processing methods influence the quality of fermented African Locust bean (Iru/ogiri/dadawa) *Parkia biglobosa*. J App Sc. Res. 2010; 6(11):1656-1661.
- 4. Ojewumi ME, Omoleye JA, Ajayi AA. Optimum Fermentation Temperature for the Protein Yield of *Parkia biglobosa* Seeds Iyere. Proceeding of the 3<sup>rd</sup> International conference on African Development Issues CUICAD. 2016a; 584-587. Ota, Ogun-state: Nigeria. ISSN 2449-075X.
- Ojewumi ME, Adesanmi AA, Obanla OM, Awolu OO, Ojewumi EO. Pozzolanic properties of Waste Agricultural Biomass - African Locust Bean Pod Waste. World Journal of Environmental Biosciences, 2016b; 6(3):1-7.
- 6. Campbell-Platt G. African locust bean *Parkia* species and its West African fermented food products, Dawadawa. Eco Food Nutr J, 1980; 9:123-132.
- Ouoba LII, Rechinger KB, Barkholt V, Diawara B, Traore AS, Jakobsen M et al. Degradation of proteins during the fermentation of African locust bean Parkia biglobosa by strains of Bacillus subtilis and Bacillus pumilus for Soumbala. J App Micro, 2003; 94:396-402.
- 8. Antai SP, Ibrahim MH. Microorganisms associated with African locust Bean *Parkia filicoidea* Welw fermentation for dawadawa production. J App Bact, 1986; 61:145-148.
- 9. Popoola TOS, Akueshi CO. Microorganisms associated with the fermentation of soybean for the production of daddawa a condiment. Nig Food J, 1985; 2:194-196.
- 10. Gberikon GM, Ameh JB, Ado SA, Umoh VJ.

- Physicochemical properties of powdered condiments of *P. biglobosa* fermented with and without mixed *Bacillus species* A and B as inocula and subjected under different drying conditions. J App Sc. Res. 2014; 6(1):115-120.
- 11. Soetan KO, Akinrinde AS, Adisa SB. Comparative Studies on the Proximate Composition, Mineral and Anti-nutritional Factors in the Seeds and Leaves of African Locust Bean *Parkia biglobosa*. Annals of Food Sc. & Tech. 2014; 15(1):70-74.
- 12. Ojewumi ME, Omoleye JA, Ajayi AA. The Effect of Starter Culture on the Protein Content in Fermented African Locust Bean Parkia biglobosa Seeds. International Journal of Engineering and Technology. 2016c; 5(4):249-255.
- Nwokeleme CO, Obeta J, Ugwuanyi O. Evolution of Volatile Flavour Compounds during Fermentation of African Oil Bean *Pentaclethra macrophylla* Benth Seeds for Ugba Production. International Journal of Food Science, 2015; Article ID 706328, 8 pages. doi:10.1155/2015/706328.
- 14. Odebunmi EO, Oluwaniyi OO, Bashiru MO. Comparative Proximate Analysis of Some Food Condiments. J App Sc. Res, 2009; 6:272-274.
- 15. Odunfa SA, Adesomoju A. Fatty acid composition of African locust beans *Parkia biglobosa*. Chemistry Microbiology Technology Lebesm, 1986; 10:125-127.
- 16. AOAC. Official Method of Analysis. 17th Ed. Gaithersburg, USA: Association of Official Analytical Chemists, 2000.
- 17. Sarkar PK, Tamang JP. Changes in the microbial profile and proximate composition during and controlled fermentations of soybeans. Food Microbiology, 1995; 12:317-325.
- 18. Omafuvbe BO, Shonukan O, Abiose S. Microbiological and biochemical changes in the traditional fermentation of soybean for soy-daddawa-a Nigerian food condiment. Journal of Food Microbiology, 2000; 17:469-474.
- 19. Omodara TR, Aderibigbe EY. Effects of the starter culture on the quality of fermented Parkia biglobosa. International Journal of Bio-Technology. 2013; (3)4:33-40.
- Omafuvbe BO, Olumuyiwa S, Falade BA, Osuntogun S, Adewusi RA. Chemical and Biolochemical changes in African Locust Beans Parkia biglobosa and Melon Citrullus vulgaris seeds during fermentation to condiments. Pakistan Journal of Nutrition. 2004; 3(3):140-145.
- 21. Esenwah CN, Ikenebomeh MJ. Processing effects on the nutritional and anti-nutritional contents of African locust bean Parkia biglobosa Benth Seed. Pakistan Journal Nutrition. 2008; 7(2):214-217.
- 22. Abdoulaye O. Influence of process condition on the digestibility of African Locust Bean (Parkia biglobosa) Starch. American J Food Technol, 2012; 7:552-561.
- 23. Ojewumi ME. Optimizing the Conditions and Processes for the Production of Protein Nutrient from Parkia biglobosa Seeds. PhD Thesis submitted to Covenant University in partial fulfillment for the award of Doctorate degree in Chemical Engineering, Ota, Nigeria, 2016d.