

Contents lists available at ScienceDirect

MethodsX

journal homepage: www.elsevier.com/locate/mex

Method Article

Functionalization of La_{0.33}Ca_{0.67}MnO₃ with biologically active small ligand at room temperature



A. Edobor-Osoh^{a,*}, P. de la Presa^c, B.I. Ita^b, K.O. Ajanaku^a, F.E. Owolabi^a, S.J. Olorunshola^d

^a Department of Chemistry, Covenant University, Ota, Ogun State, Nigeria

^b Department of Pure and Applied Chemistry, Calabar, Cross River State, Nigeria

^c Instituto de Magnetismo Applicado, UCM-ADIF-CSIS, Madrid, Spain

^d Department of Biological Sciences, Covenant University, Ogun State, Nigeria

ABSTRACT

We report the structural, morphological and optical activities of a paramagnetic manganite ($La_{0.33}Ca_{0.67}MnO_3$) synthesized at 900 °C. A simple method of formation of complex was employed. A complex was formed between a pre-prepared manganite dissolved in HCl and reacted with an organic ligand (ethyl 4-nitrobenzoate). The optical and antimicrobial properties of a complex were determined. The Ultraviolet-visible and Fourier-transform infra-red spectroscopy were used in monitoring optical activities of the resulting product. It was observed to absorb in the visible region (205 nm and 256 nm). The peaks observed from the infra-red spectra indicated that the reaction occurred at the nitroso end of the ethyl 4nitrobenzoate. The bacterial inhibitory property of the LCMO-ethyl 4-nitrobenzoate was determined against Pseudomonas aeruginosa, Candida albican, Aspergillus niger and Staphylococcus auerus. It was observed to inhibit the growth of all the microbes with zone of inhibitions of 60 mm, 56 mm, 45 mm and 32 mm, for Pseudomonas aeruginosa, Candida albican, Aspergillus niger and Staphylococcus auerus, respectively.

- The method used was simple and a complex was formed within 6 h without the use of complicated equipment.
- The method requires no heat treatment and can be prepared at room temperature.
- LCMO-ethyl 4-nitrobenzoate was biologically active against *Pseudomonas aeruginosa,Candida albican, Aspergillus niger* and *Staphylococcus aureus.*

© 2019 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY license (http:// creativecommons.org/licenses/by/4.0/).

A R T I C L E I N F O Method name: Functionalization of manganite Keywords: Functionalization, Manganite, Ethyl 4-nitrobenzoate, Nanoparticles, Antimicrobial agent Article history: Received 15 August 2018; Accepted 5 March 2019; Available online 22 March 2019

E-mail address: abiola.edobor@covenantuniversity.edu.ng (A. Edobor-Osoh).

https://doi.org/10.1016/j.mex.2019.03.004

^{*} Corresponding author at: Department of Chemistry, Covenant University, Km. 10, Idiroko road, P. M. B. 1023, Ota, Ogun State, Nigeria.

^{2215-0161/© 2019} The Authors. Published by Elsevier B.V. This is an open access article under the CC BY license (http:// creativecommons.org/licenses/by/4.0/).

I	
Subject Area:	Materials Science
More specific subject area:	Biomedical nanoparticles
Method name:	Functionalization of manganite
Name and reference of	1. Giri, A., Makhal, A., Ghosh, B., Ravchaudhuri, A. K. and Pal, S. K. (2014) Functionalization
original method:	of manganite nanoparticles and their interaction with biologically relevant small ligand:
	Picosecond time-resolved FRET studies. Nanoscale, 2, 2704-2709
	2. Iniama, G., de la Presa, P., Alonso, J. M., Multigner, M., Ita, B. I., Cortés-Gil, R., Ruiz-
	González, M. L., Hernando, A., and Gonzalez-Calbet, J. M. (2014). Unexpected ferromagnetic
	ordering enhancement with crystallite size growth observed in $La_{0.5}Ca_{0.5}MnO_3$
	nanoparticles. Journal of Applied Physics, 116 (11,390)
Resource availability:	NA

Specifications Table

Method details

Background

Perovskites are compound with a general formula ABO₃ (A are rare earth metal (lanthanides), B sites have transition metal (Fe, Mn, Cr, etc.); manganites are perovskites with transition element manganese (Mn) as a component of the compound, thereby modifying the general formula to AMnO₃. Doping of manganites at the A-site with a divalent metal confers interesting intrinsic properties such as magneto-caloric effect, colossal magneto-resistivity, and transport properties [1–9]. These properties have been studied intensely by a number of researchers. Although a lot is known about the structural and morphological properties of the surface of the solid state, however, the nature of the surface of metal oxides in the presence of an aqueous solution has not really understood [10,11]. It is assumed that at pH < 6, the properties of the surface area of the metal does not change, therefore making it possible to functionalize the surface of the metal by modification, whereas at pH > 6 the manganite dissolves into different components; pyrolusite ($MnO_2(s)$) and manganese ions $Mn^{2+}(aq)$. The complex formed between the dissolved manganite and ethyl 4- nitrobenzoate indicates that the aqueous manganese react readily with the organic ligand. Although a lot is known on the antimicrobial properties of nanoparticle [12]; however, biological activities of manganites generally have been studied by only a few researchers [13,14]. In recent times the stability of dextran coated manganite (La_{0.7}Sr_{0.3}MnO₃ (LSMO)) in water and phosphate buffer solution was determined. It was noted that the stability of dextran coated $La_{0.3}Ca_{0.7}MnO_3$ were better than the bare manganite. It was also observed that the specific absorption rate of the dextran coated LSMO was significant and the manganites were not toxic to the L929 cells when introduced [15].

Reagents used in the study

The reagents used in this study included:

Hydrochloric acid (1 M), concentrated nitric acid, lanthanum oxide, calcium carbonate, manganese carbonate, ethylene glycol, citric acid monohydrate, ethyl 4-nitrobenzoate

Equipment used in the study

The equipments used in analyzing the sample are as follows: Scanning Electron Microscope (JEOL JSM 6400) X-ray Diffraction (PANalytical X'pert Pro MPD diffractometre) Magnetic stirrer (Stuart Agar Hot plate/Magnetic stirrer Model Number: AGG3788C) Autoclave (Thermo-Fischer precision compact oven) UV-visible Spectrophotometer (Genesys 30 visible spectrophotometre) Fourier Transform Infra-red Spectrophotometre (Perkin-Elmer λ25 and a Thermo Nicolet FTIR

200 spectrophotometre)

Preparation of the samples (Synthesis of the manganite)

- 1 In preparing the sample, analytical grade La₂O₃, MnCO₃ and CaCO₃ were stoichiometrically reacted forming a greenish solution, based on method used by [16].
- 2 La_2O_3 was pre-prepared by heating in a furnace for 2 h at 900 °C to remove the water of crystallization embedded in the sample.
- 3 5 mL of nitric acid was heated up to 50 $^\circ\text{C}$ on a hot plate.
- 4 1.36 g of lanthanum acid was added into the acidic solution on the hot plate until it turned transparent.
- 5 10 g of citric acid was added to the solution.
- 6 1.68 g of MnCO₃ was added to the solution and stirred until the solution became transparent.
- 7 1.68 g of CaCO₃ was added into the solution and it was allowed to dissolve completely.
- 8 10 mL of ethylene was added as a fuel for combustion.
- 9 The solution was heated until it evaporated to a xerogel.
- 10 With continuous heating of the xerogel the sample combusted into brown powder.
- 11 The brown powder was milled for 2 h using agate mortar.
- 12 The milled sample was heated up for 24 h at 600 °C to remove all organic compounds in the sample.
- 13 The sample was sintered at 900 °C for 2 h.
- 14 The structural and morphological properties of the sample were determined using XRD and SEM, respectively.

The proposed equation of reaction based on the precursors used is as described below:

$$\frac{1}{6}La_2O_3 + \frac{2}{3}CaCO_3 + MnCO_3 \to La_{0.33}Ca_{0.67}MnO_3 + CO_2 \uparrow$$
(1)

Morphological properties of La_{0.33}Ca_{0.67}MnO₃

The morphological property of the manganite was determined by using JEOL JSM 6400 Scanning Electron Microscopy (SEM).

- 1 The samples analyzed by SEM imaging were dry and clean.
- 2 The oil on the surface of the sample holder was cleaned up before use.
- 3 A small portion of the sample was taken and gently sprinkled using a small pipette on a carbon tape which was mounted on a sample holder.
- 4 Blower was used to remove loosely held samples.
- 5 The samples were coating with a conductive carbon metal (graphene), using a sputter coater to reduce the charging effect before SEM images are taken [17,18].

The surface topography as well as the sizing of the sample was done via SEM. It was determined to be heterogeneously shaped and sized. The sample was observed to be porous as shown in Fig. 1. The nominal composition $La_{0.32}Ca_{0.68}Mn_{0.97}O_3$ was determined for the sample. The slight difference between the nominal and experimental composition was probably due unavoidable errors incurred during synthesis.

Structural properties of the sample

X-ray diffractometre (XRD) of the sample at room temperature was obtained using Cu-K_{α} radiation of wavelength 1.54056 Å in a PANalytical X'pert Pro MPD diffractometre.

- 1 A minute volume of the sample was tapped onto a glass slide for analysis. The sample was loaded into the XRD instrument and installed by opening the door of the instrument.
- 2 A quick scan of the structural properties of the sample was done at 10,000–100,000 scan range at 0.030 step size. The mean crystallite size **<D**> was calculated from the Scherer's formula.



Fig. 1. SEM micrograph of LCMO 900.

3 Fullprof programme based on Rietveld method [19] was used for phase analysis. This gave the cell dimensions, atomic position, R factor and a model diffraction pattern utilizing PCR file code 56712.

Fig. 2 shows the XRD diffractograph of $La_{0.33}Ca_{0.67}Mno_3$ broadening of the peaks in Fig. 2 indicates that the sample was nanoscaled. It was also observed that the sample was a single perovskite phase in the orthorhombic, Pnma crystal structure.

The crystallite size was determined by using the Scherer's Eq. (2):

$$D = \frac{0.9\lambda}{\beta_{FWHM} \text{Cos}\theta}$$

(2)

Where, $\lambda = 0.154 \text{ nm}$

 β_{FWHM} = Full width at half maximum of the X-ray reflection θ = Position of the highest intensity

Surface functionalization of La_{0.33}Ca_{0.67}MnO₃ manganite oxide

The samples were synthesized based on a modified version of methods used by [20].

- 1 Pre-prepared La_{0.33}Ca_{0.67}MnO₃ was dissolved in 150 mL 1 M HCl with pH 5.6 and stirred homogenously for an hour at room temperature, a greenish yellow solution was observed.
- 2 The ethyl 4-nitobenzoate was used as purchased from Merck, Germany.
- 3 The ethyl 4-nitrobenzoate was dissolved in 150 mL of ethanol.
- 4 150 mL of the ligand solution was divided into five aliquots of 30 mL each.
- 5 30 mL of the sample was added into 150 mL of the manganite solution while stirring at room temperature for at least 2 h.
- 6 The synthesis of the complex was monitored using thin layer chromatography (TLC) plates and UVvis spectrophotometer.
- 7 After this, the beaker containing the solution was covered with perforated aluminum foil and left undisturbed for two weeks for the crystals to grow.
- 8 The recovered crystal was filtered and dried.
- 9 The sample was weighed to obtain the percentage yield.
- 10 Optical analysis of the sample was done by using Fourier-transform Infra-red and Ultravioletvisible spectrophotometer. Fig. 3 and Table 1 describe the Fourier Transform infra red peaks. The Ultra-violet peak assignment was done in Table 2. It was utilized to determine the site of reaction between the metal and the ligand.



Fig. 2. XRD Pattern of La_{0.33}Ca_{0.67}MnO₃ oxide annealed at 900 °C.

Antimicrobial analysis of the LCMO-ethyl 4-nitrobenzoate Determination of the zone of inhibition

- 1 Prior to the analysis the sample was dissolved in a universal solvent (Dimethyl sulphoxide) (DMSO) and the negative control was done.
- 2 The zone of inhibition (Table 3) and of the dissolved samples was determined by using the well diffusion method as described by [21].
- 3 Nutrient agar was used to sub-culture the microorganisms.
- 4 The agar was heated so as to convert it to liquid state after which it was cooled to 50 °C in a water bath.
- 5 Streaks of the *Staphylococcus aureus* (ATCC 25923), *Pseudomonas aeruginosa* (ATCC 1544), *Aspergillus niger* (cultured) and *Candida albican* (ATCC 10231) was put into the agar and left alone to set.
- 6 The petri dish was put in an incubator for 24 h at 37 °C.



cm⁻¹

Fig. 3. FT-IR image of the LCMO-Ethyl 4-nitrobenzoate complex showing the peak.

Table 1

The Infra-red peaks observed for the complex formed and the organic ligand (ethyl 4-nitrobenzoate).

Functional Groups	Range (cm ⁻¹) LCMO-Ethyl 4-nitrobenzoate	Range (cm ⁻¹) Ethyl 4-nitrobenzoate
C ₂ H ₅ COO (Ester)	1714.50	1712.00
Ar—C=O	1603.65	1607.00
Ar-N=0	1526.15	1527.00
Mn-O	476.63	-

Table 2

Comparison of the ultraviolet-visible wavelength (nm) of ethyl 4-nitrobenzoate and LCMO-ethyl 4-nitrobenzoate.

S/N	Name of Sample	Wavelength (nm) (log ϵ_{max})	
1 2	Ethyl 4-nitrobenzoate ICMO+Ethyl 4-nitrobenzoate	256 (5.065) 256 (5.421)	202 (4.512) 205 (4.972)
-	Eenio · Ethyr i introbenzoute	250 (5:121)	205 (1.572)

Table 3

The zone of inhibition of the La_{0.33}Ca_{0.67}MnO₃ –ethyl 4-nitrobenzoate (complex), controls anti-fungal (nitrofuratoin) and antibacterial (gentamicin).

Sample	Concentration	Staphylococcus aureus (ATCC 25923) (mm)	Aspergillus niger (cultured) (mm)	P. aeruginosa (ATCC 15442) (mm)	Candida albican (ATCC 10231) (mm)
La _{0.33} Ca _{0.67} MnO ₃ – Ethyl 4-nitrobenzoate (Complex)	1000 mg/mL	32	45	60	56
Nitrofuratoin (control)	300 µg/mL	-	32	-	28
Gentamicin (control)	10 μg/disc	25	-	24	-

- 7 After this period, holes of about 5 mm to the edge of the plates were drilled into the petri dishes by using a sterile cock borer.
- 8 200 μ l of the dissolved solution was inoculated into the perforated well using a micro pipette.
- 9 The petri dish was pre-diffused for 30 min and kept in an incubator for 24 h at 37 °C, until an obvious reduction of the potency of the complex was observed.
- 10 The zone of inhibition of the sample was determined in mm.
- 11 10 µg/disc of a standard anti-fungal drug (nitrofuratoin) and an antibacterial drug (gentamicin) were used as controls.

Determination of the minimum inhibition concentration (MIC)

- 1 The minimum inhibitory concentration (MIC) of the samples were determined using the broth dilution method as shown in Table 4.
- 2 The antibiotic (LCMO-ethyl 4-nitrobenzoate) stock solution was prepared by dissolving in DMSO.
- 3 Sterile test tubes were used in conducting the test.
- 4 The antibiotic was diluted in the range of 0-300 mg/mL.
- 5 A final volume of 1 mL of the dilution was used in the MIC test.
- 6 Broth suspension of the bacteria was selected from plates that have been incubated for 24 h at 35 °C.
- 7 1 mL of the diluted antibiotics was added and a positive control tube containing the broth and the microbes was also used
- 8 The inoculated tubes were incubated for 24 h at 35 °C.
- 9 The turbidity of the tubes was determined visually.

Table 4

The minimum inhibitory concentration of the $La_{0.33}Ca_{0.67}\text{MnO}_3$ – ethyl 4-nitrobenzoate (complex).

Micro-organism	Minimum inhibitory concentration (mg/mL)
Candida albican	31.25
Staphylococcus aureus	7.80
Pseudonomas aeruginosa	7.81

- 10 The growth of the microbes in the tubes containing the antimicrobial agent was compared with the amount of growth with no antimicrobials.
- 11 The lowest concentration that inhibited the growth of the microbes was considered as the minimum inhibitory concentration.

Additional information

This method of synthesis was utilized by [20] using a cyclo-mixer for the capped La_{0.67}Sr_{0.33}MnO₃citrate. The capped sample was reacted with 4-nitrophenylanthranilate (NPA). In our study the same method was utilized by using a magnetic stirrer. The optical property of the sample was determined using ultraviolet-visible and Fourier-Transform infra-red (FT-IR) spectrophotometer showed that a complex was formed between the metal and the organic ligand. HCl was used to dissolve the sample at a pH > 6 and the resulting manganese formed a complex with a small organic ligand (ethyl 4nitrobenzoate). The slight bathochromic shift in the band position 202 nm indicates the coordination reaction, but the optical activity of the product was attributed to the organic ligand [22]. The antimicrobial property of La_{0.33}Ca_{0.67}MnO₃-ethyl 4-nitrobenzoate was determined by using *Candida albican*, *Pseudomonas aeruginosa*, *Aspergillus niger* and *Staphylococcus aureus*.

Acknowledgements

We would like to acknowledge Covenant University, Ota, Ogun State for all their support in this research. We would also like to thank Instituto de Magnetismo Applicado, UCM-ADIF-CSIS, Madrid, Spain for allowing the use of their equipment for this research.

References

- A. Rostamnejadi, M. Venkatesan, P. Kameli, H. Salamati, J.M.D. Coey, Magnetocaloric effect in La_{0.67}Sr_{0.33}MnO₃ manganite above room temperature, J. Magn. Magn. Mater. 323 (2011) 2214–2218.
- [2] L.I. Koroleva, D.M. Zashchirinskii, A.S. Morozov, R. Symczak, Magnetocaloric effect in manganite, J. Exp. Theor. Phys. 115 (4) (2012) 653–661.
- [3] F.E.N. Ramirez, F.F. Ferreira, W.A. Alves, J.F.Q. Rey, J.A. Souza, Magnetic, structural, and transport properties at very high temperature in manganites, J. Magn. Magn. Mater. 324 (24) (2012) 2011–2018.
- [4] H. Zhao, L. Hu, Z. Zhang, The effect of reducing time on the magnetoresistance of manganite La_{0.67}Sr_{0.02}Cu_{0.10} 0.03 MnO₃ at a temperature of 30 °C, J. Nanomater. 2012 (2012) 1–5, doi:http://dx.doi.org/10.1155/2012/426037.
- [5] A.G. Gamzatov, A.B. Batdalov, G.G. Ramazanova, L.G. Nevruzova, Weak amplification of the magnetocaloric effect in manganites, Phase Trans. 87 (3) (2013) 305–311, doi:http://dx.doi.org/10.1080/01411594.2013.826802.
- [6] M. Baldini, T. Muramatsub, M. Sherafatic, H.-K. Maob, L. Malavasie, P. Postorinog, S. Satpathyh, V.V. Struzhkinb, Origin of colossal magnetoresistance in LaMnO₃ manganite, Proc. Natl. Acad. Sci. U. S. A. 112 (35) (2015) 10869–10872.
- [7] J. Jeon, J. Jung, K.H. Chow, Electron beam induced tunneling magnetoresistance in spatially confined manganite, Nanoscale 9 (48) (2017) 19304–19309.
- [8] Z. Zhang, J. Li, W. Zhou, C. Yang, Q. Cao, D. Wang, Y. Du, Mechanism of enhancement in magnetoresistance properties of manganite perovskite ceramics by current annealing, Ceram. Int. 44 (4) (2018) 3760–3764.
- [9] Y. Wang, Y. Zhu, H. Liu, H. Lin, T. Miao, Y. Yu, H. Furong, W. Wang, J. Sun, L. Yin, J. Shen, A large enhancement of magnetocaloric effect by chemical ordering in manganites, J. Mater. Chem. C 6 (2018) 1224–1228.
- [10] C.J. Matocha, E.J. Elzings, D.L. Sparks, Reactivity of Pb(II) at the Mn(III, IV) (oxyhyr)oxide-water interface, Environ. Sci. Technol. 35 (14) (2001) 2967–2972.
- [11] M. Ramstedt, B.M. Andersson, A. Shchukarev, S. Sjöberg, Surface properties of hydrous manganite (γ-MnOOH): a potentiometric, electroacoustic, and X-ray photoelectron spectroscopy study, Langmuir 20 (19) (2004) 8224–8229.
- [12] A.A. Akinsiku, E.O. Dare, K.O. Ajanaku, J.A. Adekoya, S.O. Alayande, A.O. Adeyemi, Synthesis of silver nanoparticles by plantmediated green method: optical and biological properties, Bionanoscience 10 (2016) 17180.

- [13] C.O. Ehi-Eromosele, J.A.O. Olugbuyiro, A. Edobor-Osoh, A.A. Adebisi, O.A. Bamgboye, J. Ojeifo, Magneto-structural and antimicrobial properties of sodium doped lanthanum manganite magnetic nanoparticles for biomedical applications: influence of silica coatings, J. Biomim. Biomater. Biomed. Eng. 7 (2296-9845) (2018) 117–127.
- [14] D. De, S.M. Mandal, S.S. Gauri, R. Bhattacharya, S. Ram, S.K. Roy, Antibacterial effect of lanthanum calcium manganite (La_{0.67}Ca_{0.33}MnO₃) nanoparticles against *Pseudomonas aeruginosa* (ATCC 27853), J. Biomed. Nanotechnol. 6 (2010) 1–7.
- [15] N.D. Thorat, R. Patil, V.M. Khot, A.B. Salankhe, A.I. Prasad, K.C. Barick, R. Ningthojam, S.H. Pawar, Highly water -dispersible surface –functionalized LSMO nanoparticles for magnetic hyperthermia application, New J. Chem. 39 (9) (2013) 2733.
- [16] G. Iniama, P. de la Presa, J.M. Alonso, M. Multigner, B.I. Ita, R. Cortés-Gil, M.L. Ruiz-González, A. Hernando, J.M. Gonzalez-Calbet, Unexpected ferromagnetic ordering enhancement with crystallite size growth observed in La_{0.5}Ca_{0.5}MnO₃ nanoparticles, J. Appl. Phys. 116 (11) (2014) 113901.
- [17] K. Jores, W. Mehnert, M. Drecusler, H. Bunyes, C. Johan, K. MAder, Investigation on the stricter of solid lipid nanopartuicles and oil-loaded solid nanoparticles by photon correlation spectroscopy, field flow fractionation and transmission electron microscopy, J. Control. Release 17 (217-) (2004) 227.
- [18] S.L. Pal, U. Jana, P.K. Manna, G.P. Mohanta, R. Manavalan, Nanoparticles: an overview of preparation and characterization, J. Appl. Pharm. Sci. 01 (06) (2011) 228–234.
- [19] H.M. Rietveld, A profile refinement method for nuclear and magnetic structures, J. Appl. Crystallogr. (65) (1969) 2 part 2.
- [20] A. Giri, A. Makhal, B. Ghosh, A.K. Ravchaudhuri, S.K. Pal, Functionalization of manganite nanoparticles and their interaction with biologically relevant small ligand: pico-second time-resolved FRET studies, Nanoscale 2 (2014) 2704–2709.
- [21] R.V. Geetha, R. Anitha, In-vitro evaluation of antibacterial activity three herbal methicilin resistant *Staphylococcus aureus* (MRA), J. Pharm. Sci. Res. 5 (10) (2013) 207–209.
- [22] S. Gao, L. Li, I. Vohra, D. Zha, L. You, Differential metal-binding properties of dynamic acylhyrazone polymers and their sensing applications, R. Soc. Open Sci. 4 (8) (2017) 170466, doi:http://dx.doi.org/10.1098/rsos.170466L.