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## Evaluation of Methods for the Analysis of Untreated and Processed Lignocellulosic Biomasses

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Chapter

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## Abstract

The overall efficiency of the transformation of lignocellulosic materials to usable products as chemicals and fuels must be governed by adequate analysis of products before and after treatments. Using some promising technologies, lignocelluloses which are biomasses from marine plant and trees, grains, food and non-food crops, and wood-based can give products as fuel alcohol and other chemicals. Various methods of transformation from feedstock to valuable end products are discussed in the scientific literature. Therefore, yields must justify methods used for biomass transformations. As a result, adequate compositional analysis of these processing stages is needed. In this chapter, standard common methods such as gravimetric, chromatography, spectroscopic and their variations for analysis on both untreated and treated lignocelluloses are highlighted. The ease of the use and challenges with recommendations to their applicability to quantifying lignocelluloses fractionations for reproducibility and to be representative are discussed. With biomass technology, virtually all and even more products that can be produced from fossil energy can also be produced from biomass energy. Adequate analysis is therefore necessary.

## Keywords

Biomass Analysis Pretreatment techniques Compositional analysis Chemical products

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## References

1. 1.

U.P. Agarwal, 1064 nm FT-Raman spectroscopy for investigations of plant cell walls and other biomass materials. *Front. Plant Sci.* **5**, 1–12 (2014) [CrossRefGoogle Scholar](#)

2. 2.

U.P. Agarwal, Raman Spectroscopy in the analysis of cellulose nanomaterials, in *Nanocelluloses: Their Preparation, Properties and Applications*, ed. by U. P. Agarwal, R. H. Atalla, A. Isogai, (American Chemical Society, Washington, DC, 2017), pp. 75–90 [CrossRefGoogle Scholar](#)

3. 3.

U.P. Agarwal, Analysis of cellulose and lignocellulose materials by Raman Spectroscopy: A Review of the current status. *Molecules* **24**, 1–16 (2019) [CrossRefGoogle Scholar](#)

4. 4.

H. Amiri, K. Karimi, Improvement of acetone, butanol, and ethanol production from woody biomass by using organosolv pretreatment. *Bioprocess Biosyst. Eng.* **38**, 1959–1972 (2015) [CrossRefGoogle Scholar](#)

5. 5.

C.S. Anna, W. de Souza, Microscopy as a tool to follow deconstruction of lignocellulosic biomass, in *Current Microscopy Contribution to Advances in Science and Technology*, ed. by A. Mendez-Vilas, (FORMATEX, 2012), pp. 639–645 [Google Scholar](#)

6. 6.

D.S. Argyropoulos, Quantitative phosphorus-31 NMR analysis of lignin, a new tool for the lignin chemist. *J. Wood Chem. Technol.* **14**, 45–63 (1994) [Google Scholar](#)

7. 7.

A.A. Awosusi, A. Ayeni, R. Adeleke, M.O. Daramola, Biocompositional and thermodecompositional analysis of South African agro-waste corncob and husk towards production of biocommodities. *Asia Pac. J. Chem. Eng.* **12**, 960–968 (2017) [CrossRefGoogle Scholar](#)

8. 8.

A.O. Ayeni, O.A. Adeeyo, O.M. Oresegun, T.E. Oladimeji, Compositional analysis of lignocellulosic materials: Evaluation of an economically viable method suitable for woody and non-woody biomass. *Am. J. Eng. Res.* **4**, 14–19 (2015) [Google Scholar](#)

9. 9.

A.O. Ayeni, M.O. Daramola, Lignocellulosic biomass waste beneficiation: Evaluation of oxidative and non-oxidative pretreatment methodologies of South African corn cob. *J. Environ. Chem. Eng.* **5**, 1771–1779 (2017) [CrossRefGoogle Scholar](#)

10. 10.

A.O. Ayeni, M.O. Daramola, A. Awoyomi, et al., Morphological modification of *Chromolaena odorata* cellulosic biomass using alkaline peroxide oxidation pretreatment methodology and its enzymatic conversion to biobased products. *Cogent Eng.* **5**, 1509665 (2018) [CrossRefGoogle Scholar](#)

11. 11.

A.O. Ayeni, M.O. Daramola, P.T. Sekoai, et al., Statistical modelling and optimization of alkaline peroxide oxidation pretreatment process on rice husk cellulosic biomass to enhance enzymatic convertibility and fermentation to ethanol. *Cellulose* **25**, 2487–2504 (2018) [CrossRefGoogle Scholar](#)

12. 12.

A.O. Ayeni, F.K. Hymore, S.N. Mudliar, et al., Hydrogen peroxide and lime based oxidative pretreatment of wood waste to enhance enzymatic hydrolysis for a biorefinery: Process parameters optimization using response surface methodology. *Fuel* **106**, 187–194 (2013)[CrossRef](#)[Google Scholar](#)

13. 13.

A.O. Ayeni, R. Ogu, A.A. Awosusi, M.O. Daramola, Alkaline peroxide oxidation pretreatment of corn cob and rice husks for bioconversion into bio-commodities: Part A-Enzymatic convertibility of pretreated rice husks to reducing sugar. Paper presented at the 24th European Biomass Conference and Exhibition, Amsterdam, Netherlands, 6–9 June 2016 (2016)[Google Scholar](#)

14. 14.

A.O. Ayeni, J.A. Omoleye, F.K. Hymore, et al., Effective alkaline peroxide oxidation pretreatment of shea-tree sawdust for the production of biofuels: Kinetics of delignification and enzymatic conversion to sugar and subsequent production of ethanol by fermentation using *Saccharomyces cerevisiae*. *Braz. J. Chem. Eng.* **33**, 33–45 (2016)[CrossRef](#)[Google Scholar](#)

15. 15.

A.B. Bjerre, A. Plöger, T. Simonsen et al., Quantification of solubilized hemicellulose from pretreated lignocellulose by acid hydrolysis and high-performance liquid chromatography, RISØ National Laboratory, Roskilde, Denmark, pp. 1–39 (1996)[Google Scholar](#)

16. 16.

C.D. Blasi, G. Signorelli, C. Di Russo, G. Rea, Product distribution from pyrolysis of wood and agricultural residues. *Ind. Eng. Chem. Res.* **38**, 2216–2224 (1999)[CrossRef](#)[Google Scholar](#)

17. 17.

N. Carpita, M. McCann, The cell wall, in *Biochemistry and Molecular Biology of Plants*, ed. by B. Buchanan, W. Gruissem, R. Jones, (American Society of Plant Physiologists, Rockville, 2000), pp. 52–108[Google Scholar](#)

18. 18.

F. Carrilo, X. Colom, J.J. Suñol, et al., Structural FTIR analysis and the thermal characterization of lyocell and viscose-type fibers. *Eur. Polym. J.* **40**, 2229–2234 (2004)[CrossRef](#)[Google Scholar](#)

19. 19.

D. Ciolacu, F. Ciolacu, V.I. Popa, Amorphous cellulose-Structure and characterization. *Cellul. Chem. Technol.* **45**, 13–21 (2011)[Google Scholar](#)

20.20.

S.C. Corgié, H.M. Smith, L.P. Walker, Enzymatic transformations of cellulose assessed by quantitative high-throughput fourier transform infrared spectroscopy (QHT-FTIR). *Biotechnol. Bioeng.* **108**, 1509–1520 (2011)[CrossRefGoogle Scholar](#)

21.21.

R.C.N.R. Corrales, M.F.M. Teixeira, C.C. Perrone, et al., Structural evaluation of sugar cane bagasse steam pretreated in the presence of CO<sub>2</sub> and SO<sub>2</sub>. *Biotechnol. Biofuels* **5**, 36 (2012)[CrossRefGoogle Scholar](#)

22.22.

F.B. de Oliveira, J. Bras, M.T. Borges Pimenta, et al., Production of cellulose nanocrystals from sugar cane bagasse fibers and pith. *Ind. Crop. Prod.* **93**, 48–57 (2016)[CrossRefGoogle Scholar](#)

23.23.

Z. Dische, Colour reactions of pentoses, in *Methods in Carbohydrate Chemistry*, ed. by R. L. Whistler, M. L. Wolfrom, (Academic Press, New York, 1962), pp. 475–514[Google Scholar](#)

24.24.

B.S. Donohoe, T.B. Vinzant, R.T. Elander, V.R. Pallapolu, Y.Y. Lee, R.J. Garlock, V. Balan, B.E. Dale, Y. Kim, N.S. Mosier, M.R. Ladisch, M. Falls, M.T. Holtzapple, R. Sierra-Ramirez, J. Shi, M.A. Ebrik, T. Redmond, B. Yang, C.E. Wyman, B. Hames, S. Thomas, R.E. Warner, Surface and ultrastructural characterization of raw and pretreated switchgrass. *Bioresour. Technol.* **102**, 11097–11104 (2011)[CrossRefGoogle Scholar](#)

25.25.

A. Duval, M. Lawoko, A review on lignin-based polymeric, micro- and nano structured materials. *React. Funct. Polym.* **83**, 78–96 (2014)[CrossRefGoogle Scholar](#)

26.26.

M. Fan, D. Dai, B. Huang, Fourier transform infrared spectroscopy for natural fibres, in *Fourier Transform – Materials Analysis*, ed. by S. Mohammed Salih, (InTech, 2012). <https://doi.org/10.5772/35482>

27.27.

V. Ferreira-Leitão, C.C. Perrone, J. Rodrigues, et al., An approach to the utilisation of CO<sub>2</sub> as impregnating agent in steam pretreatment of sugar cane bagasse and leaves for ethanol production. *Biotechnol. Biofuels* **3**, 1–8 (2010) [CrossRef](#) [Google Scholar](#)

28.28.

T. Foyle, L. Jennings, P. Mulcahy, Compositional analysis of lignocellulosic materials: Evaluation of methods used for sugar analysis of waste paper and straw. *Bioresour. Technol.* **98**, 3026–3036 (2007) [CrossRef](#) [Google Scholar](#)

29.29.

J. Fromm, B. Rockel, S. Lautner, Lignin distribution in wood cell walls determined by TEM and backscattered SEM techniques. *J. Struct. Biol.* **143**, 77–84 (2003) [CrossRef](#) [Google Scholar](#)

30.30.

S.C. Fry, *The Growing Cell Wall: Chemical and Metabolic Analysis* (Blackburn Press, New Jersey, 2000), pp. 1–333 [Google Scholar](#)

31.31.

B. Godin, R. Agneessens, P. Gerin, J. Delcarte, Structural Carbohydrates in a plant biomass: Correlations between the detergent fiber and dietary fiber methods. *J. Agric. Food Chem.* **62**, 5609–5616 (2014) [CrossRef](#) [Google Scholar](#)

32.32.

B. Godin, R. Agneessens, P. Gerin, et al., Composition of structural carbohydrates in biomass: Precision of a liquid chromatography method using a neutral detergent extraction and a charged aerosol detector. *Talanta* **85**, 2014–2026 (2011) [CrossRef](#) [Google Scholar](#)

33.33.

B. Godin, R. Agneessens, S. Gofflot, et al., Revue sur les méthodes de caractérisation des polysaccharides structuraux des biomasses lignocellulosiques. *Biotechnol. Agron. Soc. Environ.* **15**, 165–182 (2011) [Google Scholar](#)

34.34.

B. Godin, S. Lamaudière, R. Agneessens, et al., Chemical composition and biofuel potentials of a wide diversity of plant biomasses. *Energy Fuel* **27**, 2588–2598 (2013) [CrossRef](#) [Google Scholar](#)

35.35.

A.A. Guilherme, P.V.F. Dantas, E.S. Santos, et al., Evaluation of composition, characterization and enzymatic hydrolysis of pretreated sugar cane bagasse. *Braz. J. Chem. Eng.* **32**, 32–33 (2015) [CrossRefGoogle Scholar](#)

36.36.

H.M. Hernández-Hernández, J.J. Chanona-Pérez, E. Terrés, A. Vega, P. Ligerero, R.R. Farrera-Rebollo, S. Villanueva, Microscopy and spectroscopy tools for the description of delignification. *Cell. Chem. Technol.* **53**, 87–97 (2019) [CrossRefGoogle Scholar](#)

37.37.

S.E. Jacobsen, C.E. Wyman, Cellulose and hemicellulose hydrolysis models for application to current and novel pretreatment processes. *Appl. Biochem. Biotechnol.* **84–86**, 81–96 (2000) [CrossRefGoogle Scholar](#)

38.38.

H. Janshekar, A. Fiechter, Lignin: Biosynthesis, application and biodegradation. *Adv. Biochem. Biotechnol.* **27**, 119–178 (1983) [Google Scholar](#)

39.39.

J.P. Joseleau, J. Comtat, K. Ruel, Chemical structure of xylans and their interaction in the plant cell walls, in *Progress in Biotechnology – Xylans and Xylanases*, ed. by J. Visser, (Elsevier, 1992), pp. 1–15 [Google Scholar](#)

40.40.

X. Ju, M. Bowden, E.E. Brown, X. Zhang, An improved X-ray diffraction method for cellulose crystallinity measurement. *Carbohydr. Polym.* **123**, 476–481 (2015) [CrossRefGoogle Scholar](#)

41.41.

H.-J. Jung, Analysis of forage fiber and cell walls in ruminant nutrition. *J. Nutr.* **127**, 810S–813S (1997) [CrossRefGoogle Scholar](#)

42.42.

H.-J. Jung, J. Lamb, Prediction of cell wall polysaccharide and lignin concentrations of alfalfa stems from detergent fiber analysis. *Biomass Bioenergy* **27**, 365–373 (2004) [CrossRefGoogle Scholar](#)

43.43.

K. Karimi, M.J. Taherzadeh, A critical review of analytical methods in pretreatment of lignocelluloses: Composition, imaging, and crystallinity. *Bioresour. Technol.* **200**, 1008–1018 (2016) [CrossRefGoogle Scholar](#)

44.44.

E.M. Karp, B.S. Donohoe, M.H. O'Brien, et al., Alkaline pretreatment of corn stover: Bench-scale fractionation and stream characterization. *ACS Sustain. Chem. Eng.* **2**, 1481–1491 (2014)[CrossRefGoogle Scholar](#)

45.45.

K.L. Larsen, S. Barsberg, Environmental effects on the lignin model monomer, vanillyl alcohol, studied by Raman spectroscopy. *J. Phys. Chem. B* **115**, 11470–11480 (2011)[CrossRefGoogle Scholar](#)

46.46.

S. Li, S. Xu, S. Liu, C. Yang, et al., Fast pyrolysis of biomass in free-fall reactor for hydrogen-rich gas. *Fuel Process. Technol.* **85**, 1201–1211 (2004)[CrossRefGoogle Scholar](#)

47.47.

L. Liu, X.P. Ye, A.R. Womac, S. Sokhansanj, Variability of biomass chemical composition and rapid analysis using FT-NIR techniques. *Carbohydr. Polym.* **81**, 820–829 (2010)[CrossRefGoogle Scholar](#)

48.48.

Y. Lu, Y.-C. Lu, H.-Q. Hu, F.-J. Xie, X.-Y. Wei, X. Fan, Structural characterization of lignin and its degradation products with spectroscopic methods: Review. *J. Spectrosc.*, 1–15 (2017)[Google Scholar](#)

49.49.

J.S. Lupoi, E. Gjersing, M.F. Davis, Evaluating lignocellulosic biomass, its derivatives, and downstream products with Raman spectroscopy: A Review. *Front. Bioeng. Biotechnol.* **3**, 1–18 (2015)[CrossRefGoogle Scholar](#)

50.50.

J.S. Lupoi, S. Singh, R. Parthasarathi, B.A. Simmons, R.J. Henry, Recent innovations in analytical methods for the qualitative and quantitative assessment of lignin. *Renew. Sust. Energ. Rev.* **49**, 871–906 (2015)[CrossRefGoogle Scholar](#)

51.51.

J.S. Lupoi, S. Singh, B.A. Simmons, R.J. Henry, Assessment of lignocellulosic biomass using analytical spectroscopy: An evolution to high-throughput techniques. *Bioenergy Res.* **7**, 1–23 (2014)[CrossRefGoogle Scholar](#)

52.52.



J.S. Lupoi, E.A. Smith, Characterization of woody and herbaceous biomasses lignin composition with 1064 nm dispersive multichannel Raman spectroscopy. *Appl. Spectrosc.* **66**, 903–910 (2012)[CrossRefGoogle Scholar](#)

53.53.

M. Makarem, C.M. Lee, K. Kafle, S. Huang, I. Chae, H. Yang, J.D. Kubicki, S.H. Kim, Probing cellulose structures with vibrational spectroscopy. *Cellulose* **26**, 35–79 (2019)[CrossRefGoogle Scholar](#)

54.54.

T. Mani, P. Murugan, J. Abedi, et al., Pyrolysis of wheat straw in a thermogravimetric analyser: Effect of particle size and heating rate on devolatilization and estimation of global kinetics. *Chem. Eng. Res. Des.* **88**, 952–958 (2010)[CrossRefGoogle Scholar](#)

55.55.

D.A. Matos, I.P. Whitney, M.J. Harrington, et al., Cell walls and the developmental anatomy of the *Brachypodium distachyon* stem internode. *PLoS One* **8**(11), e80640 (2013). <https://doi.org/10.1371/journalCrossRefGoogle Scholar>

56.56.

R.L. McCreery, *Raman Spectroscopy for Chemical Analysis* (Wiley, New York, 2000), p. 448[CrossRefGoogle Scholar](#)

57.57.

M.W. Meyer, J.S. Lupoi, E.A. Smith, 1064 nm dispersive multichannel Raman spectroscopy for the analysis of plant lignin. *Anal. Chim. Acta* **706**, 164–170 (2011)[CrossRefGoogle Scholar](#)

58.58.

G.L. Miller, Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Anal. Chem.* **31**, 426–428 (1959)[CrossRefGoogle Scholar](#)

59.59.

T. Milne, H. Clum, F. Agblevor, D.K. Johnson, Standard analytical methods. *Biomass Bioenergy* **2**, 341–366 (1992)[CrossRefGoogle Scholar](#)

60.60.

B.I. Na, S.J. Chang, K.H. Lee, G. Lee, J.W. Lee, Characterization of cell wall structure in dilute acidpretreated biomass by confocal Raman microscopy and enzymatic hydrolysis. *Biomass Bioenergy* **93**, 33–37 (2016)[CrossRefGoogle Scholar](#)

61.61.

M.L. Nelson, R.T. O'Connor, Relation of certain infrared bands to cellulose crystallinity and crystal lattice type. Part II. A new infrared ratio for estimation of crystallinity in cellulose I and II. *J. Appl. Polym. Sci.* **8**, 1325–1341 (1964)[CrossRefGoogle Scholar](#)

62.62.

D.R. Nhuchhen, P. Abdul Salam, Estimation of higher heating value of biomass from proximate analysis: A new approach. *Fuel* **99**, 55–63 (2012)[CrossRefGoogle Scholar](#)

63.63.

R.T. O'Connor, E.F. DuPré, D. Mitcham, Application of infrared absorption spectroscopy to investigations of cotton and modified cottons. Part I: Physical and crystalline modifications and oxidation. *Text. Res. J.* **28**, 382–392 (1958)[CrossRefGoogle Scholar](#)

64.64.

S.Y. Oh, D.I. Yoo, Y. Shin, et al., FTIR analysis of cellulose treated with sodium hydroxide and carbon dioxide. *Carbohydr. Res.* **340**, 417–428 (2005)[CrossRefGoogle Scholar](#)

65.65.

J. Parikh, S.A. Channiwala, G.K. Ghosal, A correlation for calculating HHV from proximate analysis of solid fuels. *Fuel* **84**, 487–494 (2005)[CrossRefGoogle Scholar](#)

66.66.

S. Park, J.O. Baker, M.E. Himmel, P.A. Parilla, D.K. Johnson, Cellulose crystallinity index: Measurement techniques and their impact on interpreting cellulase performance. *Biotechnol. Biofuels* **3**, 1754–6834 (2010)[CrossRefGoogle Scholar](#)

67.67.

M. Poletto, H.L. Ornaghi Júnior, A.J. Zattera, Native cellulose: Structure: Characterization and thermal properties. *Materials* **7**, 6105–6119 (2014)[CrossRefGoogle Scholar](#)

68.68.

Y. Pu, B. Hallac, A.J. Ragauskas, Plant biomass characterization: Application of solution- and solid-state NMR spectroscopy, in *Aqueous Pretreatment of Plant Biomass for Biological and Chemical Conversion to Fuels and Chemicals*, (Wiley, 2013), pp. 369–390[Google Scholar](#)

69.69.

J. Puls, J. Schusiel, Chemistry of hemicelluloses: Relationship between hemicellulose structure and enzymes required for hydrolysis, in *Hemicelluloses and Hemicellulases*, ed. by M. P. Coughlan, G. P. Hazlewood, (Portland Press, 1993), pp. 1–27 [Google Scholar](#)

70.70.

C.A. Rezende, P. Maziero, E.R. Azevedo, et al., Chemical and morphological characterization of sugarcane bagasse submitted to delignification process for enhanced enzymatic digestibility. *Biotechnol. Biofuels* **4**, 54 (2011) [CrossRefGoogle Scholar](#)

71.71.

D.R. Robert, G. Brumow, Quantitative estimation of hydroxyl group in milled wood lignin from spruce and in a dehydrogenation polymer from coniferyl alcohol using  $^{13}\text{C}$  NMR spectroscopy. *Holzforschung* **38**, 85–90 (1984) [CrossRefGoogle Scholar](#)

72.72.

L. Segal, J.J. Creely, A.E. Martin, et al., An empirical method for estimating the degree of crystallinity of native cellulose using the X-ray diffractometer. *Text. Res. J.* **29**, 786–794 (1959) [CrossRefGoogle Scholar](#)

73.73.

M.J. Selig, T.B. Vinzant, M.E. Himmel, et al., The effect of lignin removal by alkaline peroxide pretreatment on the susceptibility of corn stover to purified cellulolytic and xylanolytic Enzymes. *Appl. Biochem. Biotechnol.* **155**, 397–406 (2009) [CrossRefGoogle Scholar](#)

74.74.

C. Sheng, J.L.T. Azevedo, Estimating the higher heating value of biomass fuels from basic analysis data. *Biomass Bioenergy* **28**, 499–507 (2005) [CrossRefGoogle Scholar](#)

75.75.

Y.D. Singh, P. Mahanta, U. Bora, Comprehensive characterization of lignocellulosic biomass through proximate, ultimate and compositional analysis for bioenergy production. *Renew. Energy* **103**, 490–500 (2017) [CrossRefGoogle Scholar](#)

76.76.

A. Sluiter, B. Hames, R. Ruiz et al., Determination of sugars, byproducts, and degradation products in Liquid Fraction Process Samples, National Energy Renewable Laboratory, Golden Colorado, USA, NREL/TP-510-42623 (2008a) [Google Scholar](#)

77.77.

A. Sluiter, B. Hames, R. Ruiz et al., Determination of structural carbohydrates and lignin in biomass, National Renewable Energy Laboratory, Golden, CO, USA, pp. 1–16, (2008b) [Google Scholar](#)

78.78.

A. Sluiter, R. Ruiz, C. Scarlata et al., Determination of extractives in biomass. National Energy Renewable Laboratory, Golden, CO, USA, NREL/TP-510-42619, pp. 1–9, (2008c) [Google Scholar](#)

79.79.

J. Sluiter, R. Ruiz, C. Scarlata, et al., Compositional analysis of lignocellulosic feedstocks. 1. Review and description of methods. *J. Agric. Food Chem.* **58**, 9043–9053 (2010) [CrossRef](#) [Google Scholar](#)

80.80.

S. Tao, S. Khanizadeh, H. Zhang, et al., Anatomy, ultrastructure and lignin distribution of stone cells in two *Pyrus* species. *Plant Sci.* **176**, 413–419 (2009) [CrossRef](#) [Google Scholar](#)

81.81.

N. Terashima, M. Yoshida, J. Hafrén, et al., Proposed supramolecular structure of lignin in softwood tracheid compound middle lamella regions. *Holzforschung* **66**, 907–915 (2012) [CrossRef](#) [Google Scholar](#)

82.82.

N. Terinte, R. Ibbett, K.C. Schuster, Overview on native cellulose and microcrystalline cellulose I structure studied by X-ray diffraction (waxd): Comparison between measurement techniques. *Lenzinger Ber.* **89**, 118–131 (2011) [Google Scholar](#)

83.83.

O. Theander, P. Aman, E. Westerlund, et al., Total dietary fiber determined as neutral sugar residues uronic acid residues, and Klason lignin (the Uppsala method): Collaborative study. *J. AOAC Int.* **78**, 1030–1044 (1995) [CrossRef](#) [Google Scholar](#)

84.84.

O. Theander, E. Westerlund, Quantitative analysis of cell wall components, in *Forage Cell Wall Structure and Digestibility*, ed. by H. Jung, D. Buxton, R. Hatfield, J. Ralph, (American Society of Agronomy, Madison, WI, 1993), pp. 83–104 [Google Scholar](#)

85.85.

B. Uner, I. Karaman, H. Tanriverdi, D. Ozdemir, Determination of lignin and extractive content of Turkish Pine (*Pinus brutia* Ten.) trees using near infrared spectroscopy and multivariate calibration. *Wood Sci. Technol.* **45**, 121–134 (2011)[CrossRef](#)[Google Scholar](#)

86.86.

P.J. Van Soest, Collaborative study of acid-detergent fiber and lignin. *J. Assoc. Off. Anal. Chem.* **56**, 781–784 (1973)[Google Scholar](#)

87.87.

P.J. Van Soest, R.H. Wine, Use of detergents in the analysis of fibrous feeds. IV. Determination of plant cell-wall constituents. *J. Assoc. Off. Anal. Chem.* **50**, 50–55 (1967)[Google Scholar](#)

88.88.

J.H. Wiley, R.H. Atalla, Band assignments in the Raman spectra of celluloses. *Carbohydr. Res.* **160**, 113–129 (1987)[CrossRef](#)[Google Scholar](#)

89.89.

S. Yao, G. Wu, M. Xing, S. Zhou, J. Pu, Determination of lignin content in *Acacia* spp. using near-infrared reflectance spectroscopy. *Bioresources* **5**, 556–562 (2010)[Google Scholar](#)

90.90.

T.-F. Yeh, C. H-m, J.F. Kadla, Rapid prediction of solid wood lignin content using transmittance near-infrared spectroscopy. *J. Agric. Food Chem.* **52**, 1435–1439 (2004)[CrossRef](#)[Google Scholar](#)

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