The *Anopheles gambiae* Insecticidal Targets Made Bare by *In-silico* Analysis

^{1, 2}Marion Adebiyi, ³Olubanke Ogunlana, ^{1, 2}Ezekiel Adebiyi, ¹Segun Fatumo

¹Department of Computer and Information Sciences

²Covenant University Bioinformatics Research,

³Department of Biological Sciences (Biochemistry Unit)

Covenant University,

Ota, Nigeria

Jason L. Rasgon

Department of Entomology

Center for Infectious Diseases Dynamics and the Huck
Institutes of the Life Sciences
Pennsylvania State University, USA

Abstract—several works had attempted to use genomics to explain the mode of mosquito resistance and predict drug target. The use of insecticides in various ways has been the major malaria vector control strategy being deployed lately, mostly pyrethroid, the major recommended compound class for IRS, ITNs and LLITNs. Resistance to drugs and insecticides has continually obstructed vector/malaria control strategies. The advert effect is so enormous in the Sub-Saharan African; its socioeconomic impact is unquantifiable in every measure. Thus, the quick necessity for the development and elucidation of potent, cheap and efficient new potential insecticidal targets, especially those in the class pyrethroid for the malaria vector, A. gambiae. In this work, an updated Anopheles gambiae biochemical metabolic network (AnoCyc ver1.0), otherwise known as pathway genome database (PGDB) was extracted, the database was reconstructed by developing a computational graph model in an approach that modeled the metabolic network of the organism as a bipartite graph, deployed the concept of choke point, load point and reaction without deviation to determine the essential enzymatic reactions in the networks. Each potential drug target to their corresponding gene/protein and such encoding protein sequences were extracted. (PDB) was blasted for genes that have structure or homologue of >= 30 sequence identity. Finally, we deployed Overton and Barton Score (OB-Score) and ParCrys prediction to rank proteins by their likely success in crystallization. 61 potential insecticidal candidate targets was made bare, one clinically validated insecticidal target and others with biological evidence in the literature. Seven of these targets ideally stand out and have no homology with other vertebrates. These in depth dissection of the biochemical metabolic networks of the Anopheles effectively identified the ideal gene products and specifically extract essential enzymes as new potential insecticidal target against A. gambiae.

Index Terms—Anopheles gambiae, metabolism, insecticidal targets, in-silico, essential enzymes

I. Introduction

Insecticide resistance is an inherited characteristic involving changes in one or more insect genes [17, 18]. It is also a major public health challenge combating world efforts on malaria control. The malaria vector *Anopheles gambiae* (A. gambiae) has developed resistance to all existing classes of insecticides, particularly pyrethroids (the only class approved for Indoor Residual Spray [IRS]

and Long-Lasting Insecticide Treated Net [LLITNs]) [1]. Identification of novel insecticidal targets for the development of more effective insecticides is a critical issue [2]. However, deciding which gene products are ideal insecticidal targets remains a difficult task. In the malaria parasite Plasmodium falciparum, Fatumo et al. [3,4] developed a computational method to investigate the topology of biochemical metabolic networks to mine new viable enzymatic drug targets which some were subsequently validated experimentally, [6]. Uniquely it glutamyl-tRNA discovered that amidotransferase of P. falciparum can be inhibited by 6diazo-5-oxonorleucine. This has been confirmed by an invivo study observing P. berghei infected mice [6]. We envisaged that the dissection and comprehensive study of biochemical metabolic networks has great potential to effectively and specifically identify essential enzymes as potential insecticidal targets against A. gambiae.

The completion of the genome sequence of the malaria vector, *A. gambiae* [7] has also given opportunity to develop various methods to facilitate effective malaria control strategies. Thus, in a previous work, we used the PathoLogic program [8] to construct AnoCyc ver1.0, a pathway/genome database (PGDB) for *A. gambiae* AgamP3, using its annotated genomic sequence and other annotated information from UNIPROT and KEGG databases. The resulting first PGDB for *A. gambiae* AgamP3 was deployed under the www.bioCyc.org databases (http://biocyc.org/ANO2/organism-summary?object=ANO2). This database was downloaded and rigorously analyzed in this study.

II. MATERIALS AND METHODS

A. Reconstructing the metabolic network

We extracted the metabolic network data of *A.gambiae* AgamP3 from AnoCyc database, http://www.biocyc.org, Version 18.1). The metabolic network of *A. gambiae* AgamP3 was modeled as a bipartite graph consisting of alternating nodes and edges as described previously [3,5,6]. For completion, in brief, this network was a connected graph which was established by defining neighbors of reactions: two reactions were neighbors if a metabolite existed that were the product of one reaction and the substrate for the other.

This yielded a bipartite graph of alternating reaction and metabolic compound nodes

B. Computational analysis of the reconstructed metabolic network

We used the concept of Choke Point (CP) to analyze the structure of our metabolic network [9,10]. A reaction is a chokepoint if it consumes a unique substrate and produces a unique product, making it indispensable. Inactivating choke-points could lead to an organism's failure. The number of true predictions out of all predictions of their approach is often large and sometimes difficult to prioritized [3,4], this makes it difficult for an experimentalist to choose the appropriate potential drug target when developing inhibitors as effective therapeutics [9].

As a combinatorial approach, we adapted the concept of Reaction without Deviation (RWD) [3,4] to further analyse our network. RWD are otherwise known as essential reactions. RWD analysis identified reactions within the network that are choke-points for which there are no other routes (deviations) within the network to replace the reactions. Figure 1 [3,4] indicates that an enzymatic reaction is considered a potential drug targets when such a reaction is ONLY a choke point (CP) as well as reaction without deviation (RWD). It also confirms that the knocked out reaction is a choke-point but may not be essential for the organism if the dashed lines exist in the metabolic network. Our approach inspects the network for such deviations. Thus, the RWD analysis investigates a reaction by deleting such a reaction from the metabolic network and further checks whether a chosen product could still be produced without the deleted reaction. This method can test multiple knock-outs merged with isolating essential enzymes on a high throughput manner to predict effective drug combinations [3,5].

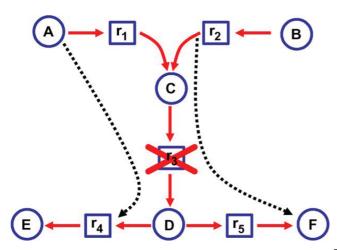


Fig. 1. The knocked out reaction is a choke-point but may not be essential for the organism if the dashed lines exist in the metabolic network. Our approach inspects the network for such deviations. (Reactions are boxes while metabolites are circles). [3,4].

III. RESULTS

Choke points analysis yielded 448 reactions, while implementing the RWD analysis yielded 645 reactions. 156 reactions were found to have their reversed copy included and were regarded as a duplicate of the forward reaction captured, therefore every duplicated reaction was discarded and we were left with 489 reactions. We compared the list of possible insecticidal target as captured by both the CP and RWD analysis, 264 reactions falls into the category of set of reactions that forms a tie (Interception) between the two methods (TABLE I.).

TABLE I. IMPORTANT STATISTICS FROM CP AND RWD ANALYSIS

Description	Quantity	
Total number of CP enzyme before sieving for	896	
duplicates		
CP Enzymes captured after sieving	448	
Total number of RWD enzymes before sieving	645	
RWD enzymes captured after sieving	489	
Interception of CP + RWD =	264	
TIE		

Each reaction on the tie (intercept) list of 264 reactions was matched to its corresponding enzymatic gene or protein. Targets that had more than one catalyzing enzymes and those without a valid corresponding enzymatic gene/protein and EC number were discarded, reducing the tie list to 173 enzymes. We extracted encoding protein sequences for each of the 173 enzymes from the Uniprot database server, where 153 out of 173 has their encoding sequences on this database. A total of 9 gene IDs were not resolvable. We again performed a comprehensive Position Specific Iteration alignment search on PDB database to determine if any of the protein sequences has a structure or homologue removal of 30% PDB sequence identity cut off. All potential targets with >30% identity to protein in PDB were discarded. In total 128 proteins on the target list had a <=30% identity to proteins in PDB. A PSI-BLASTp of every sequence against the Human, Bird (Chicken) and Fish (Tilapia) genome databases was performed to eliminate targets with homology to non-target organisms. Finally, OB & ParCrys [13,14], prediction was used to rank the 128 protein sequences according to their estimated likely success in crystallization. OB & ParCrys estimates a protein's propensity to produce diffraction quality crystals and its predictions are intended to provide guidance in selecting best targets (Table 2). The top 61 proteins on the predicted targets list classified as high scoring were selected as our final list of predicted insecticidal targets. We discarded the others.

TABLE II. OB & PARCRYS RATING TABLE FOR PREDICTED TARGETS LIST

OB & ParCrys Ranking	No of Genes in Class
High scoring	61
Amenable	33
Recalcitrant	31
Irresolvable	3
Total	128

TABLE 1. of the Appendix detailed the complete list of these 61 predicted insecticidal targets.

Next we compared these 61predicted enzymes with the gold standard list of 20 experimentally validated targets as represented on table 3 below. Those highlighted in red color are the enzymes that were not found on our reconstructed network. The rest of the enzymes were found on our network and 5 of them were captured as CP and / or RWD enzymes.

TABLE II. of the appendix summarizes the results of their homology tests with protein transcripts of the Human, Bird (Chicken) and Fish (Tilapia) genomes. Those highlighted in green have no significant homology (all E-values > 0.01) with Human and any other of the two organisms. See Tables 7, 8 and 9 of the supplementary material for full details of these homology tests.

A. The Gold Standard List

ECN

The gold standard list is a collection of proposed drug/ insecticidal targets from literature. On scanning a variety of established databases of insecticidal bank and literatures, we gathered a total of 20 reactions contained on our network. Of the 20 list, 11 were not found on our network. We extracted from various literature, 3 targets of clinically proven insecticides and the remaining 9 were proposed insecticidal targets, with proposal based on biological validation/ evidence via in vitro growth inhibition of the A. gambiae. To equally compare all predictions with the gold standard, every network on the network was mapped to its corresponding enzyme classification (EC) number. To measure the performance estimation, reactions without EC number were discarded. The complete list of the gold standard is given in TABLE III.

TABLE III. THE GOLD STANDARD LIST

EC Number	REACTION	References			
1.1.1.145	3-beta —hydroxysteroid dehydrogenase/Delta 5 >4-isomerase	Curr Med Chem. 2008			
1.4.3.4	Amine oxidase [flavin containing] B	Emerging drugs in neuropathic pain. Expert Opin Emerg Drugs. 2007			
1.13.11.27	4-hydroxyphenylpyruvate dioxygenase	Expert Opin Emerg Drugs. 2009 Jun			
1.14.15.6	Cytochrome P450 11A1, mitochondrial	Nucleic Acids Res. 2008 Jan			
2.4.1.34	1,3-Beta-Glucan synthase	Mini Rev Med Chem. 2007			
3.1.1.7	Acetylcholine esterase	J Med Chem. 2005 Feb 10			
2.3.1.178	Diaminobutanoate acteyltransferase	Peters et al., 1990			
2.6.1.46	Diaminobutyrate pyruvate transaminase.	Vandenende et al., 2004			
2.6.1.96	4-aminobutyrate— pyruvate transaminase	Van Cauwenberghe, O.R. and Shelp, B.J, 1999			
1.3.7.1	6-hydroxynicotinate reductase.	Alhapel et al., 2006			
1.17.1.5	Nicotinate dehydrogenase.	Yang et al., 2010			
1.17.2.1	Nicotinate dehydrogenase	Yang et al., 2009			
1.17.3.3	6-hydroxynicotinate	Wieser et al., 1997			

	dehydrogenase	
2.4.2.11	Nicotinate	Hara et al., 2007
	phosphoribosyltransferase	
2.7.1.173	Nicotinate riboside kinase	Tempel et al., 2007,
		plos comp.
4.2.1.23	Spinosyn A	Tempel et al., 2007,
		plos comp
4.1.4.12	Tetraethylammonium-	Tempel et al., 2007,
	r(TEA)	plos comp
4.1.14.7	Bungarotoxin (α7-	Tempel et al., 2007,
	nAChR)	plos comp
1.1.11.141	Gamma-	San Framcisco
	glutamyltransferase-rxn	inclusion, 2007
1.1.1.42	poly(oxy-1,2-	San Framcisco
	ethanediyl),alphaisodecyl-	inclusion, 2007.
	omegahydroxy-	
	phosphate	

IV. CONCLUSION

Using these methods we computationally predicted list of 61 insecticidal targets for *A. gambiae*. This includes one with clinical validation. Out of these we identified 7 targets with no significant homology to Human, tilapia or chicken. For the known 20 experimental validated targets, out of the 9 found on our reconstructed network, 5 were detected by our computational analysis. Table 1 of the appendix elucidates the details of the 61 predicted insecticidal targets and information available for each of them from literature.

It is also clear from our analysis of the PGDB databases and data [19] that there is need for further rigorous pursuit of the manual and automatic curation of the biochemical metabolic network for *A. gambiae*.

Our analysis of the *P. falciparum* network has revealed that curation helped to close gaps and link up dangling ends. Thus, Following the results in our work on *P. falciparum* in [3,4,6], and on *A. gambiae* in [19], we derived insecticidal targets in *A. gambiae* with no significant homology to Human and also to Bird (Chicken) and Fish (Tilapia). These can be confirmed on the appendix tables and supplementary document/ data.

ACKNOWLEDGMENTS

MA was supported by the Covenant University Staff Development Program. We thank Adaobi Okafor (a research assistant at Covenant University bioinformatics unit) for her contribution.

REFERENCES

- [1] Ranson H, Claudianos C, Ortelli F, Abgrall C, Hemingway J, Sharakhova MV, Unger MF, Collins FH, Feyereisen R: Evolution of supergene families associated with insecticide resistance. *Science* 2002, 298(5591):179-181.
- [2] Hemingway J, Beaty BJ, Rowland M, Scott TW, Sharp BL. Trends Parasitol 22: 308-312, 2006.
- [3] Fatumo, S., Plaisma, K., Mallm, J-P., Schramm, G., Adebiyi, E., Oswald, M. Eils, R. and Koenig, R. (2009), "Estimating novel potential drug targets of Plasmodium falciparum by analysing the metabolic network of knock-out strains in silico", Elsevier Journal of Infection, Genetics and Evolution, Vol. 9, No. 3, pp.351-358.
- [4] Fatumo, S., Adebiyi, E., Schramm, G., Eils, R. and Koenig, R. (2009), "An in silico approach to design an efficient malaria drug to combat the malaria resistance problem", IACSIT, IEEE Computer Society Press, Vol. 17, pp. 564-569.

- [5] Fatumo, S., Kitiporn, P., Adebiyi, E. and Koenig, R. Elsevier Journal of Infection, Genetics and Evolution, 2010 Sep 7 (Epub ahead of print)
- [6] Kitiporn Plaimas, Yulin Wang, Solomon O. Rotimi, Grace Olasehinde, Segun Fatumo, Michael Lanzer, Ezekiel Adebiyi, Rainer König. Computational and experimental analysis identified 6-diazo-5-oxonorleucine as a potential agent for treating infection by Plasmodium falciparum. In press, 2013.
- [7] Holt R. et al., (2002), "The Genome Sequence of the Malaria Mosquito Anopheles gambiae", Journal of Science, Vol. 298, pp.129 – 149.
- [8] Karp PD, Paley S, Romero P. The Pathway Tools software. Bioinformatics. 2002;18 Suppl 1:S225-32.
- [9] Yeh, I., Hanekamp, T., Tsoka, S., Karp, P.D., Altman, R.B., 2004. Computational analysis of Plasmodium falciparum metabolism: organizing genomic information to facilitate drug discovery. Genome Res. 14, 917–924.
- [10] Rahman, S.A., Schomburg, D., 2006. Observing local and global properties of metabolic pathways: 'load points' and 'choke points' in the metabolic networks. Bioinformatics 22, 1767–1774.
- [11] Vinayagam A, del Val C, Schubert F, Eils R, Glatting KH, Suhai S, Koenig R.GOPET: a tool for automated predictions of Gene Ontology terms.BMC Bioinformatics. 2006 Mar 20;7:161.
- [12] del Val C, Ernst P, Falkenhahn M, Fladerer C, Glatting KH, Suhai S, Hotz-Wagenblatt A.ProtSweep, 2Dsweep and DomainSweep: protein analysis suite at DKFZ.Nucleic Acids Res. 2007 Jul;35(Web Server issue):W444-50. Epub 2007 May 25.
- [13] Overton & Barton (2006) "A normalised scale for structural genomics target ranking: The OB-Score." FEBS Lett. 580, 4005-4009
- [14] Overton, Padovani, Girolami & Barton (2008) "ParCrys: A Parzen Window Density Estimation Approach to Protein Crystallisation Propensity Prediction." Bioinformatics 24:901-907.
- [15] Caspi R, Altman T, Dale JM, Dreher K, Fulcher CA, Gilham F, Kaipa P, Karthikeyan AS, Kothari A, Krummenacker M, Latendresse M, Mueller LA, Paley S, Popescu L, Pujar A, Shearer AG, Zhang P, Karp PD. (2010) "The MetaCyc database of metabolic pathways and enzymes and the BioCyc collection of pathway/genome databases," Nucleic Acids Research 38:D473-9.
- [16] Hemingway, J., Field, L. and Vontas, G. (2002). "An Overview of Insecticide Resistance", Journal of Science, Vol. 298, pp. 96 - 97.
- [17] Hemingway, J. (2002). "The Molecular Basis of Two Contrasting Metabolic Mechanisms of Insecticide Resistance". Insect Biochem Mol Biol, Vol. 30, pp. 1009 – 101.
- [18] Adebiyi Marion, Segun Fatumo, Jason Rasgon, Ezekiel Adebiyi. (2015). "Computational Analysis of Anopheles gambiae Metabolism to Facilitate Insecticidal Target Discovery" (Under review)

APPENDIXES

TABLE IV. THE 61 PREDICTED INSECTICIDAL TARGETS AND INFORMATION AVAILABLE FOR EACH FROM LITERATURE

NAME	EC	GENE ID	Remarks	Reference	PDB
	1.1.1.145		May be involved in		
3-beta-hydroxy-delta5-steroid		AGAP0059	pheromones/hormones synthesis.	Simard J. et al.,	
dehydrogenase		84	May be a good target.	2005.	4DQ
	1.1.1.35		Involved Fat metabolism, lipid		
3-hydroxyacyl-coa		AGAP0077	degradation/metabolism (May not	Hillmer and	
dehydrogenase Peroxid-rxn	1 11 1 7	84	be a strong target)	Gottschalk, 1974	4B3F
Peroxid-rxn	1.11.1.7		(A good target) involved in oxidation of toxic reductants.		
			response to environmental		
			stresses such as wounding,		
		AGAP0003	pathogen attack and oxidative		
		96	stress	Rapoport et al., 1994	4G2I
	1.14.13.8		Detoxifying enzyme. Organisms		-
			have several mechanisms for		
			detoxification.		
Flavin-containing		AGAP0104	Its inhibition may not lead to		
monooxygenase		01	death directly	Uno et al., 2013	4A9
Cytochrome P450, family 3,	1.14.14.1	AGAP0122	V	I C C# 1000	
subfamily A	1.14.14.1	95	Very good target Very good target (Oxidizes	J. G Scott, 1999	3S79
	1.14.14.1		Very good target (Oxidizes xenobiotics and steroids,		
			Participates in the metabolism of		
			an as-yet-unknown biologically		
Cytochrome P450, family 3,		AGAP0122	active molecule that is a		
subfamily A		94	participant in eye development)	K. G Scott, 1999	3V81
Ecdysone-20-monooxygenase-	1.14.99.2		Good target. Involved in		-
rxn	2		synthesis of hormone that		
			controls molting. Insects		
			generally depend on molting for		
		A C A D0024	growth.	G 14 4 1 1070	
		AGAP0024 29	NB: It may take longer time to kill the insect.	Smith et al., 1979; 1983	3N9Y
Deoxyhypusine-	1.14.99.2	2)	A good target. Essential for	1703	- 31N9
monooxygenase-rxn	9		organism's viability and plays a		
			role in a wide number of		
			important processes such as cell		
			growth and proliferation regulates		
		AGAP0021	induction of autophagy and	Ober & Hartmann,	
		29	protein synthesis.	1999	
Aldehyde dehydrogenase	1.2.1.3	AGAP0036 52	Involved in cellular metabolic	Yoshida et al., 1998	45.03
(NAD+) Aldehyde dehydrogenase	1.2.1.3	AGAP0035	process Involved in cellular metabolic	Yoshida et al.,	4E3X
(NAD+)	1.2.1.3	78	process m centual metabolic	1998	
Dihydlipoxn-rxn	1.8.1.4	1,0	Process	Lamirande et al.,	-
				1993;	
			Good target. Involved in	http://enzyme.expa	
		AGAP0116	hyperactivation of spermatazoa	sy.org/EC/1.8.1.4	
		29	during capacitation and so on.		4EQ
Hexaprenyldihydroxybenzoate	2.1.1.114	AGAP0105	Catalyzes the ubiquinone	Marbois et al.,	
methyltransferase	01116	37	biosynthetic	1994	4HTI
Charing NI and the 14 and Comme	2.1.1.162	AGAP0021	III-lim arm for ation		2007
Glycine N-methyltransferase	211166	98	Unknown function	http://www.chem.q	. 3THI
	2.1.1.166			mul.ac.uk/iubmb/e	
23S rma (uridine2552-2'-O)-		AGAP0041		nzyme/EC2/1/1/16	
methyltransferase		77	Known for catalytic activity	6.html	3DO
	2.1.1.183	† · ·		Henras et al.,	. 500
				2008;	
				http://www.ebi.ac.	
				uk/thornton-	
				srv/databases/cgi-	
18S rrna (adenine1779-				bin/enzymes/GetPa	
N6/adenine1780-N6)-dimet		AGAP0044		ge.pl?ec_number=	
hyltransferase	0.1.1.201	65	Involved in ribosome biogenesis.	2.1.1.183	4AD
2-octaprenyl-methoxy-benzoq-	2.1.1.201			http://www.genom	
meth-rxn		AGAP0104		e.jp/dbget- bin/www bget?ec:	
		88	Ubiquinone biosynthesis	2.1.1.201	23.40
		1 00	Obiquinone biosynthesis	<u>2.1.1.2U1</u>	3MG

Trna (guanine57-N1/adenine58-N1)- methyltransfe rase 2.1.1.221 Trna (guanine9-N1)- methyltransferase 2.1.1.33 Trna (guanine-n7 methyltransferase 2.1.1.33 AGAP0003 tRNA processing (May be a slow target) Trna (guanine-n7 methyltransferase 2.1.1.33 AGAP00047 tRNA processing (May be a slow target) 2.1.1.35 AGAP0008 tRNA processing (May be a slow target)
Trna (guanine9-N1)- methyltransferase Trna-guanine-n7 methyltransferase-rxn AGAP0047 AGAP0047 Trna-guanine-n7 methyltransferase-rxn AGAP0047 Cap-specific mma (nucleoside- 2'-O-)-methyltra 2.1.1.62 AGAP008 AGAP008 AGAP008 AGAP008 Trna-guanine-n7 methyltransferase-rxn AGAP008 Cap-specific mma (nucleoside- 2'-O-)-methyltra 2.1.1.62 AGAP008 AGAP008 AGAP008 AGAP008 AGAP008 AGAP0090
Trma (guanine9-N1)- methyltransferase Trma-guanine-n7 methyltransferase-rxn 2.1.1.33 AGAP0047 AGAP0047 Example of the protein-glutamine gamma-glutamyltransferase AGAP0080 AGAP0090 Britip://www.genome.jp/dbget-bin/www.genome.gp/dbget-bin/www.genome.gp/dbget-bin/www.genome.gp/dbget-bin/www.genome.gp/dbget-bin/www.genome.gp/dbget-bin/www.genome.gp/dbget-bin/www.g
Mrna (2'-O-methyladenosine-N6-)-methyltransferase 2.3.2.13 AGAP0098 2.3.2.2 AGAP0099 AGAP0089 AG
AGAP0047 52 tRNA processing (May be a slow target) Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra 2.1.1.62 Mrna (2'-O-methyladenosine-N6-)-methyltransfer ase AGAP0028 tRNA processing (May be a slow target) AGAP0028 tRNA processing (May be a slow target) RNA processing (May be a slow target) AGAP0028 tRNA processing (May be a slow target) AGAP0028 tRNA processing (May be a slow target) AGAP0028 tRNA processing (May be a slow target) Protein-glutamine gamma-glutamyltransferase 2.3.2.13 AGAP0090 ps for speedy mortality. AGAP0090 ps for speedy mortality. AGAP0089 ps for speedy mortal
Cap-specific mma (nucleoside-2'-O-)-methyltra 2.1.1.62 Mrna (2'-O-methyladenosine-N6-)-methyltransfer ase AGAP0028 Protein-glutamine gamma-glutamyltransferase 2.3.2.13 AGAP0090 AGAP0080 A
Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-O-)-methyltra Cap-specific mma (nucleoside-2'-1.57 http://www.brenda - enzymes.org/php/r esult_flat.php4?ec no=2.1.1.62 Cap-specific mma (nucleoside-2'-1.57 http://www.genom e.jp/dbget-bin/www bget?ec: 2.3.2.13 Cap-specific mma (nucleoside-2'-1.57 AGAP0028 tRNA processing (May be a slow enzymes.org/php/r esult_flat.php4?ec no=2.1.1.62 Cap-specific mma (nucleoside-1-1.57 http://www.genom e.jp/dbget-bin/www bget?ec: 2.3.2.13 Cap-specific mma (nucleoside-1-1.57 AGAP0028 tRNA processing (May be a slow enzymes.org/php/r esult_flat.php4?ec no=2.1.1.62 Cap-specific mma (nucleoside-1-1.57 http://www.genom e.jp/dbget-bin/www bget?ec: 2.3.2.13 Cap-specific mma (nucleoside-1-1.57 AGAP0090 poble in transferase activities poble in transferase activities poble in transferase activities poble in transferase activities poble in transferase poble
Mrna (2'-O-methyladenosine-N6-)-methyltransfer ase AGAP0028 tRNA processing (May be a slow target)
Mrna (2'-O-methyladenosine-N6-)-methyltransfer ase AGAP0028 tRNA processing (May be a slow target)
Protein-glutamine gamma-glutamyltransferase 2.3.2.13 AGAP0090 Involved in hemolymph clot. Not sure it will be a good target for speedy mortality. AGAP0089 AGAP0089 Involved in hemolymph clot. Not sure it will be a good target bin/www_bget?ec: 2.3.2.13 AGAP0089 AGAP0089 Involved in transferase activities
Protein-glutamine gamma-glutamyltransferase AGAP0090 Not sure it will be a good target for speedy mortality. 2.3.2.13 2XZZ:A AGAP0089 AGAP0089 Gamma-glutamyltranspeptidase AGAP0089 Involved in transferase activities AGAP0089 Involved in transferase activities 2.3.2.2 3G9K:D AGAP0089 Involved in transferase activities Good target. Catalyzes a enzymes.info/php/r
Gamma-glutamyltranspeptidase AGAP0089 15 Involved in transferase activities
Gamma-glutamyltranspeptidase AGAP0089 15 Involved in transferase activities 2.5.1.1 Good target. Catalyzes a eigp/dbget-bin/www_bget?ec: 2.3.2.2 bhttp://www.brenda
Gamma-glutamyltranspeptidase 15 Involved in transferase activities 2.3.2.2 3G9K:D 2.5.1.1 http://www.brenda = cnzymes.info/php/r
Good target. Catalyzes a <u>= enzymes.info/php/r</u>
AGAP0071 precursor for several classes of esult flat.php4?ec
Farnesyl diphosphate synthase 04 essential metabolites no=2.5.1.1 PDB:2010
Gppsyn-rxn 2.5.1.1 http://www.brenda =
AGAP0045 Good target. Catalyzes a enzymes.info/php/r esult flat.php4?ec essential metabolites no=2.5.1.1 PDB:2W4
Alkylglycerone-phosphate- synthase-rxn
enzymes.org/php/r
esult_flat.php4?ec no=2.5.1.26;
AGAP0043 Brown and Snyder, 58 Lipid biosynthensis/metabolism 1982. PDB:3FW
Ceramide-kinase-rxn 2.7.1.138 http://www.brenda
AGAP0029 Unknown (Coenzyme a enzymes.org/php/r esult_flat.php4?ec no=2.7.1.138 PDB:3VZ
32 biosynthensis) no=2.7.1.138 PDB:3VZ
= enzymes.org/php/r
AGAP0100 Unknown (Coenzyme a esult flat.php4?ec no=2.7.1.33 PDB:2YX
Pyramkin-rxn 2.7.1.35 <u>http://www.brenda</u>
AGAP0059 Unknown (Coenzyme a esult flat.php4?ec
29 biosynthensis) <u>no=2.7.1.35</u> PDB:2DD
Ethanolamine-kinase-rxn 2.7.1.82
AGAP0000 enzymes.org/php/r esult_flat.php4?ec
10 (Coenzyme a biosynthensis)
Good target. Involved in many cellular processes especially enzymes.org/php/r
AGAP0069 production of lipid secondary esult flat.php4?ec
Sphingosine kinase 95 messengers. no=2.7.1.91 PDB:2JGI 2.7.7 http://www.ebi.ac.
DNA primase large subunit AGAP0063 DNA replication and other uk/interpro/entry/I PR016558 PDB:1ZZ

Gluconolact-rxn	3.1.1.17			http://www.ebi.ac.	
				uk/intenz/query?c	
		AGAP0108		md=SearchEC&ec	
	2.12.1	66	Y	=3.1.1.17	PDB:2J3E_A
	3.1.2.4			http://www.brenda	
				enzymes.org/php/r	
3-hydroxyisobutyryl-coa		AGAP0040	Involved in valine/protein	esult_flat.php4?ec	
hydrolase		97	catabolism	no=3.1.2.4	PDB:4JYL_A
N-acylneuraminate-9-	3.1.3.29			http://www.brenda	
phosphatase-rxn			Involved in a number of cellular	enzymes.org/php/r	
		AGAP0043	activity and will be a good but	esult flat.php4?ec	
		91	slow target.	no=3.1.3.29	PDB:3DDH_A
Polynucleotide-5-phosphatase-	3.1.3.33	L C A DOOG 5			
rxn		AGAP0035 17	Carbonhydrate metabolism, unsure of functions		DDD:21CA A
	3.1.3.62	17	unsure of functions	http://www.genom	PDB:3ICA_A
	0.11.0.0			e.jp/dbget-	
Multiple inositol-		AGAP0035	Involved in feeding and digestion	bin/www_bget?ec:	
polyphosphate phosphatase		55	of blood-feeding insects	3.1.3.62	PDB:1IHP_A
	3.1.3.66			http://www.genom e.jp/dbget-	
Inositol polyphosphate-4-		AGAP0001	inositol phosphate metabolic	bin/www bget?ec:	
phosphatase		24	process	3.1.3.66	PDB:1UPQ A
Ppgppsyn-rxn	3.1.7.2			www.berkeleybop.	
		AGAP0045	May be a good target. Enzyme	org/obo/MetaCyc:	
	2.2.1.22	97	involved in starvation response	PPGPPSYN-RXN	PDB:2B9C_A
	3.2.1.23			http://www.genom e.jp/dbget-	
		AGAP0020		bin/www_bget?ec:	
Beta-galactosidase		58	Carbonhydrate metabolizer.	3.2.1.23	PDB:3OG2 A
Betagalactosid-rxn	3.2.1.23			http://www.genom	-
		L G A BOOGO		e.jp/dbget-	
		AGAP0020 55	Carbonhydrate metabolizer.	bin/www_bget?ec: 3.2.1.23	DDD.1TC7 A
	3.2.1.52	33	Carbonnydrate metabonzer.	http://www.genom	PDB:1TG7_A
	3.2.1.32			e.jp/dbget-	
				bin/www_bget?ec:	
				3.2.1.52;	
				http://www.brenda	
				enzymes.info/php/r	
		AGAP0053		esult flat.php4?ec	
Beta-N-acetylhexosaminidase		81	Carbonhydrate metabolizer.	no=3.2.1.52	PDB:3SUV_A
	3.2.1.76	A C A D0105			
L-iduronidase		AGAP0105 49	May be a slow killer target.	Rome et al., 1978.	PDB:1PX8 A
Omega-amidase-rxn	3.5.1.3		1114y 00 to 210 to 1111101 tallgott	http://www.brenda	. 100.11710_11
				- ^	
				enzymes.org/php/r	
				esult_flat.php4?ec no=3.5.1.3;	
				http://www.genom	
				e.jp/dbget-	
		AGAP0126		bin/www_bget?enz	
		62	A very good target	yme+3.5.1.3	PDB:3HKX_A
	3.5.1.88	AGAP0038	A very good target. Involved in	http://enzyme.expa	
Peptide deformylase		61	protein biosynthensis	sy.org/EC/3.5.1.88	PDB:2AIA A
Allantoicase-rxn	3.5.3.4	1	Utilization of purines as	2,1019,2010.11.00	
		AGAP0045	secondary nitrogen sources, when		
		01	primary sources are limiting	Probst et al., 2005	PDB:3OYT_A
Histidine-decarboxylase-rxn	4.1.1.22	AGAP0090		Entrez Gene: histidine	
		01	Catecholamine biosynthesis	decarboxylase	PDB:20KK_A
	4.1.1.90			http://www.ebi.ac.	
				uk/intenz/query?c	
Dant gamma asalasy-1		AGAP0063	A good target (involved in blood	md=SearchEC&ec	DDD 1DZZ
Dent gamma-carboxylase Deoxyribose-p-ald-rxn	4.1.2.4	63	clotting)	=4.1.1.90	PDB:1RTS_A
Doony110030-p-aid-1AII	1.1.2.7	AGAP0091			
		61	Unknown function		PDB:3OA3_A

Myo-inositol-1-phosphate-	5.5.1.4		l] .	
synthase-rxn			Lipid and Inositol biosynt	heDonahue & Henry,	
		AGAP0118	Lipid and Phospholipid metab and Phospholipid biosynthesis.		
0 : 4 1:	(1111	30		al., 1997	PDB:3QVW_A
Serinetrna-ligase-rxn	6.1.1.11		Good target. Participates in Translation of mRNA to protein.		
			Blocking it will inhibit several		
		AGAP0082	biochemical processes especially	Berg et al., 1961	
		65	neuropeptide hormones	Deig et al., 1901	PDB:1WLE A
Aspartatetrna-ligase-rxn	6.1.1.12	03	Good target. Participates in		. FDB.TWLE_A
rispartate tina ngase imi	0.1.1.12		Translation of mRNA to protein.		
			Blocking it will inhibit several		
		AGAP0078	biochemical processes especially		
		44	neuropeptide hormones	Allen et al., 1960	PDB:4DPG A
Prolinetrna-ligase-rxn	6.1.1.15		Good target. Participates in	,	-
_			Translation of mRNA to protein.		
			Blocking it will inhibit several	Bergmann et al.,	
		AGAP0035	biochemical processes especially	1961	
		89	neuropeptide hormones		PDB:3UH0_A
Cysteinetrna-ligase-rxn	6.1.1.16		Good target. Participates in		
			Translation of mRNA to protein.		
			Blocking it will inhibit several		
		AGAP0118	biochemical processes especially	Berg et al., 1961;	
		21	neuropeptide hormones	Allen et al., 1960	PDB:2X1M_A
Phenylalaninetrna-ligase-rxn	6.1.1.20		Good target. Participates in		
			Translation of mRNA to protein.		
		AGAP0035	Blocking it will inhibit several biochemical processes especially	Berg et al., 1961;	
		17	neuropeptide hormones	Allen et al., 1960	PDB:2CXI A
Histidinetrna-ligase-rxn	6.1.1.21	17	Good target. Participates in	Alich et al., 1900	PDB.2CAI_A
Thisteanic that figure 14th	0.1.1.21		Translation of mRNA to protein.		
			Blocking it will inhibit several		
		AGAP00073	biochemical processes especially	Berg et al., 1961;	
Y	6114	5	neuropeptide hormones	Allen et al., 1960	PDB:3OD1_A
Leucinetrna-ligase-rxn	6.1.1.4		Good target. Participates in Translation of mRNA to protein.		
			Blocking it will inhibit several		
		AGAP00829	biochemical processes especially	Berg et al., 1961;	
		7	neuropeptide hormones	Allen et al., 1960	PDB:1WZ2_A
Isoleucinetrna-ligase-rxn	6.1.1.5		Good target. Participates in		
			Translation of mRNA to protein. Blocking it will inhibit several		
		AGAP00210	biochemical processes especially	Berg et al., 1961;	
		1	neuropeptide hormones	Allen et al., 1960	PDB:1QU3_A
	6.2.1.3				
Long-chain-fatty-acidcoa ligase		AGAP00859	Involved in lipid and fatty acid	Berg et al., 1961;	
ACSBG		6	metabolism	Allen et al., 1960	PDB:3PBK_A
Ubiquitinprotein-ligase-rxn	6.3.2.19		Involved majorly in cell growth and	Ardley and Robinson,	
		AGAP00557	differentiation. Will affect larvae	2005; Dou et al.,	DDD.2CNV A
	6.6.1.1	7	more. Very good pesticide target. Involved in	2012. Al-Karadaghi, 2001;	PDB:2CNX_A
	0.0.1.1	AGAP00495	bacteriochlorophyll pigment	Walker and	
Magnesium chelatase		6	biosynthesis	Weinstein, 1991	PDB:1LTL_A